

THE UNIVERSITY OF MEDICINE AND PHARMACY

"CAROL DAVILA", BUCHAREST

DOCTORAL SCHOOL

MEDICINE



*Research on the consequences of chronic ethanol impregnation in the evolution of myelic lesions in patients with spinal cord injury*

**PHD THESIS SUMMARY**

**PhD supervisor:**

**PROF.UNIV.DR. ONOSE GELU**

**PhD student:**

**DR. STOICA SIMONA-ISABELLE**

**2022**

*To my parents, with much love and admiration!*

## CONTENT

INTRODUCTION.....	13
I. GENERAL PART.....	19
1. General data on spinal cord injury.....	20
2. Chronic alcoholism and its physiopathological consequences, respectively complex, in spinal cord injury.....	23
3. Molecular biology mechanisms in spinal cord injury and data on current therapeutic possibilities.....	28
II. PERSONAL CONTRIBUTIONS.....	55
4. Working hypothesis and general objectives.....	56
5. General research methodology.....	62
6. Study 1: Retrospective data analysis.....	68
6.1. Introduction (working hypothesis and general objectives) .....	68
6.2. Material and method/ Patients and methods .....	69
6.3. Results.....	69
6.4. Discussions and partial (sectoral) conclusions.....	107
7. Study 2: Experimental evaluation of neurons from tumor cell cultures .....	109
7.1. Introduction (working hypothesis and general objectives) .....	109
7.2. Material and method/ Patients and methods .....	109
7.3. Results.....	112
7.4. Discussions and partial (sectoral) conclusions.....	144
8. Study 3: Evaluation of neural cells from primary cell cultures.....	154
8.1. Introduction (working hypothesis and general objectives) .....	154
8.2. Material and method/ Patients and method.....	154
8.3. Rezults.....	155
8.4. Discussions and partial (sectoral) conclusion.....	159
9. Conclusions and personal contributions.....	162
BIBLIOGRAPHY.....	169
ANNEXES.....	194

### List of abbreviations used in the text

>9w		Mai mult de 9 săptămâni
μL		Microlitru
μM		Micromolar
2w		2 săptămâni
ADAMTS	A disintegrin and metalloproteinase with thrombospondin motifs	Dezintegrina și metaloproteinază cu motive de trombospondină
ADN		Acid DezoxiriboNucleic
ADP	Adenosine DiPhosphat	Adenozin difosfat
AIF	Apoptosis-inducing factors	Factori inductori de apoptoză
AIM2	Absent In Melanoma 2	Absent în melanom 2
AIS	American Spinal Injury Association Impairment Scale	Scala Asociației Americane de Leziuni ale Coloanei Vertebrale
AK		Adenilat Kinază
ALDOA	ALDOlase, fructose-bisphosphate A	Aldolaza, fructozo-bisfosfat A
AMP	Adenosine Monophosphat	Adenozin monofosfat
AMPA	α Amino-3 hydroxi- 5 Metil- 4 – isoxazol Propionat	Acid α Amino-3 hidroxi- 5 Metil- 4 – isoxazol Propionic
ARE	Antioxidant Response Element	Elemente de răspuns antioxidant
AO	Acridine orange	
Apaf-1	Apoptosis protease activating factor-1	Factorul activator al proteazei apoptozei-1
ARN-m		Acidului RiboNucleic mesager
ASC	Apoptosis-associated Speck-like protein containing a CARD	Proteină cu formă de pată asociată apoptozei ce conține CARD
ATF	Activating Transcription Factor	Factorul activator al transcripției
ATP	Adenosine TriPhosphate	Adenozin trifosfat
Bcl-2	B-cell lymphoma 2	Proteinelor familiei limfomului cu celule B- 2
BDNF	Brain-Derived Neurotrophic Factor	Factorul de creștere neurotrofic derivat din creier
BDNF	Brain-Derived Neurotrophic Factor	Factorul neurotrofic derivat din creier
BH	Bcl-2 Homology	Domeniile de organizare omoloage Bcl-2
Bid	BH3-Interacting-Domain death agonist	Domeniul morții agonist interacțiunii cu BH3
BMK	Big MAPK	

BMP4	Bone morphogenetic proteins 4	Proteinele morfogenetice osoase 4
BMPR		Receptor BMP
CA9	Carbonic Anhydrase 9	Anhidraza carbonică 9
CAD	C-terminal transactivation domain	Domeniul de transactivare c-terminal
CARD	Caspase Activation and Recruitment Domain	Domeniul de activare și recrutare al caspazei
Casp	Caspase	Caspază
Caspases	Cysteinyll, aspartate-specific proteases	Proteaze cistenil-aspartat specific
CAT		Catalază
CBP	cAMP response element-binding protein-binding protein	Proteina de legare a proteinei de legare a elementului de răspuns AMPc
CD133		Prominina-1
CDH2		Caderina 2
CDK2	Ciclyn Dependent Kinases	Ciclin dependent kinaza 2
CHI3L1	Chitinase 3-like protein 1	Proteina 1 asemănătoare chitinazei 3
Cited2	Cbp/p300 interacting transactivator with Glu/Asp rich carboxy-terminal domain 2	Transactivatorul de interacțiune Cbp/p300 cu domeniul carboxy-terminal bogat în Glu/Asp 2
CO <sub>2</sub>		Dioxide de carbon
CoCl <sub>2</sub>		Clorură de cobalt
COX-2		CicloXigenza 2
CREB	cAMP response element-binding protein	Proteina de legare a elementelor de răspuns AMPc (adenozin monofosfat ciclic)
CSTB	CyStatinB	Cistatina B
CTL	C-Type Lectin	Lectinic tip C
CYLD	Cylindromatoză	Acizi grași polinesaturați
DAMP	Damage-Associated Molecular Pattern	Modele moleculare traumatice asociate
DD	Death Domain	Domenii ale morții
DFX		Deferoxamină
DISC	Death-Inducing Signalling Complex	Complexului semnalelor inductoare ale morții
Dkk	Proteinr Dickkopf	
DMEM	Dulbecco's Modified Essential Medium	Mediul de cultură
DMSO	Dimethyl Sulphoxide	Mediu de cultură celulară
EDTA		Acid etilendiaminotetraacetic
EGF-R	Epidermal-Growth Factor Receptor	Receptorul factorului de creștere epidermal
EMEM	Eagle's Minimum Essential Medium	Mediu de cultură celulară

EPO		Eritropoietină
Et-OH		Consum/ expunere etanolic
ERSR	Endoplasmic reticulum stress response	Răspunsul stresului reticulului endoplasmatic
FABPs	Fatty acid binding proteins	Familia proteinelor de legare ale acizilor grași
FADD	Fas-Associated Death Domain	Asociate domeniilor morții Fas, TRAILR1 și TRAILR2
FasL	Fas ligand	Ligand Fas
FBS		Ser fetal bovin inactivat
FDA	Fluorescein diacetate	Diacetat de fluorosceină
Fetu-B		Fetuin B
Fig.		Figură
FIH	Factor inhibiting HIF	Factorul inhibitor al HIF
Flk-1 (KDR)	Kinase insert domain receptor	Receptorul domeniului de inserție al kinazei
Flt	Fms related receptor tyrosine kinase	Tirozin kinaza receptorului fms
GABA	$\gamma$ -AminoButyric Acid	Acid Gama-AminoButiric
GDP	Guanin DiPhosphat	Guanin difosfat
GFAP	Glial fibrillary acidic protein	Proteină acidă glială
GH18)	Glycosyl hydrolases	18 glicozil hidrolazelor
Glut	Glucose transporter	Transportorul de glucoză
GPX4	Glutathione peroxidase 4	Glutation peroxidaza 4
GSDMD	Gasdermin	Gasdermină D
GSDMD-NT	Gasdermin- N Terminal domain	Domeniu N terminal al gasderminei D
GTP	Guanin TriPhosphat	Guanin trifosfat
Hes	Hairy/enhancer of split	Factor transcriptional
Hey	Hairy Ears, Y-linked	Factor transcriptional
HIF	Hypoxia Inducible Factor	Factor inducibil al hipoxiei
HMGB1	High Mobility Group Box 1	Categoria de grup cu mobilitate ridicată 1
HMOX		Hemoxigenază
Hmsc	Human Mesenchymal Stem Cells	Celule stem mezenchimale umane
Housekeeping		Gena de referință Rt-PCR
HRE	Hypoxic Response Element	Element de răspuns hipoxic
HSP	Hat Shock Protein	Proteină de șoc termic
HSR	Heat shock response	Răspunsul proteinelor de șoc termic
HTB-11 (SK-N-SH)		Linie celulară de neuroblastoma
HTB-14	Human Tumoral Brain 14	Linie celulară de glioblastom
IAP	Inhibitor of apoptosis proteins	Proteine inhibitorii ale apoptozei

IDO	Indoleamine 2,3-DiOxygenase	Indolamin-2,3-dioxigenază
IGF-1	Insulin-like Growth Factor-1	Factorul de creștere asemănător insulinei-1
IκB		Familia inhibitorilor κB
IKK	κB Inhibitor complex Kinase	Kinazei complexului inhibitor κB
IKK	κB inhibitor complex kinase	Kinaza complexului inhibitor κB
IL-		InterLeuchina -
iNOS	Inducible Nitric Oxide Synthase	Sintetazei inductibile a oxidului nitric
IRE-1a	Inositol-requiring protein-1a	Element component ERSR
JNK	c-Jun N-terminal protein Kinase	Protein kinaza c-Jun N-terminală
KCl		Clorură de potasiu
KH <sub>2</sub> PO <sub>4</sub>		Fosfat monopotasice
LAMA	Laminin	Laminină
LCN2		Lipocalina 2
LDH	Lactate DeHydrogenase	Lactat dehidrogenaza
LNGFR	Low-affinity Nerve Growth Factor Receptor	Receptorul cu afinitate scăzută pentru factorul de creștere al nervilor
LPS	Liposaccharides	Lipozaharide
LRR	Leucine Rich Repeat	repetitive bogate în leucină
LTh		Limfocite Thelper
MAPK	Mitogen-Activated Protein Kinases	Protein kinazele activate de mitogeni
miRNA	MicroRNA	MicroARN
mM		Millimolar
MMP		Matrix metaloproteaze
MOMP	Mitochondria Outer Membrane Permeabilisation	Permeabilizarea membranei mitocondriale externe
MPT	Mitochondrial Permeability Transition pore	Porilor de tranziție ai permeabilității mitocondriale
Na <sub>2</sub> HPO <sub>4</sub>		Fosfat acid de sodiu
NaCl		Clorură de sodiu
NAD	Nicotinamide-Adenine Dinucleotide	Nicotinamin dinucleotid adenină
NADPH	Nicotinamide adenine dinucleotide phosphate	Nicotinamidă adenin dinucleotidă fosfat,
NALP1	NACHT leucine-rich-repeat protein 1	Proteina repetitivă bogată în leucină NACHT 1
NCAM-L1	Neural cell adhesion molecule L1	Moleculele de adeziune neuronală L1
NCCD	Nomenclature Committee on Cell Death	Comitetul de nomenclatură pentru moartea celulelor
NCS	Neural stem cells	Celulelor stem neuronale
NEK7	NIMA-related kinase 7	Kinaza înrudită cu NIMA-7

Nemo		Modulatorul esențial al NF-kB
NeuroD1	Neuronal differentiation 1	Factor transcripțional
NFkB	Nuclear Factor Kappa-light-chain-enhancer of activated B cells	Factor nuclear kappa-amplificator de lanțuri uşoare ale celulelor B activate
NGF	Nerve Growth Factor	Factorul de creştere al nervilor
NIK		Kinaza indusă de NF-kB
NLR	The Nucleotide binding domain Leucine-rich Repeat	Domeniul nucleotidic de legare bogat în secvențe leucinice repetitive
NLRP3 (NALP3)	NLR family pyrin domain containing protein 3 (NACHT, LRR and PYD domains containing protein)	NLR și domeniul pirinic conținând Proteina 3
NLS	Nuclear localization signal	Semnalul de localizare nucleară
NMDA		N-metil D-aspartat
NOD-like receptor	Nucleotide Oligomerization Domain-like receptor	Receptori similari domeniului de oligomerizare a nucleotidelor
Non ET-OH		Fără consum/ expunere la etanol
Non TVM		Fără antecedente de TVM
Notch 1	Neurogenic locus notch homolog protein 1	Proteina omoloagă 1 a inciziei locusului neurogen
Npn		Neuropilin
Nrf2/ARE	Nuclear factor erythroid-related factor 2/ antioxidant response elements	Cale de semnalizare celulară
NT-3	NeuroTrophin-3	Neurotrofina 3
Olig2	Oligodendrocyte transcription factor 2	Factor transcriptional
OMI/ HTRA2		Familie de proteaze serinice
p38		Protein kinaza de 38 kDa
PAMP	Pathogen Associated Molecular Pattern	Modele moleculare patogene asociate
PARP	Poly (ADP-Ribose) Polymerase	Polimeraza poli-ADP riboză
PCDH		Protocaderine
PCR	Polymerase Chain Reaction	Reacției de polimerizare în lanț
PCSK1	Proprotein Convertase Subtilisin/Kexin type 1	Proprotein convertază subtilizină/ kexină tip 1
PEG		PoliEtilien Glicol
PERK	ERSR-activated protein kinase ribonucleic acid (RNA)(PKR)-like kinase	Element component ERSR
PFKL1	PhosphoFructoKinase, Liver Type 1	Fosfofructokinaza tip hepatic 1
PGK	PhosphoGlycerate Kinase	Fosfoglicerat kinaza
PHD	Prolin-hidroxilază	
PI	Propidium iodide	

PI-3K	Phosphoinositide 3-kinase	Fosfoinozitolkinazei 3
PMN		Polimorfonucleare
PON		Paraoxonaza
Pro-		Reziduu de prolină
PRR	Pattern Recognition Receptor	Modele de receptori de recunoaștere
PUFA	Polyunsaturated fatty acid chain	Acizi grași polinesaturați
pVHL	protein Von Hippel-Lindau	Proteina Von Hippel-Lindau
PYD	PYrin Domain	Domeniul pirinic
qRT-PCR	Quantitative RT-PCR	RT-PCR cantitativ
RCD	Regulated Cell Death	Moarte celulară programată/ reglementată
RE		Reticul endoplasmatic
RIP	Receptor interacting protein	Proteinelor de interacțiune cu receptorul
RLR	Retinoic acid- inducible gene (RIG)-I-Like Receptor	Similari receptorilor inductibili ai genelor acidului retinoic
RLU	Relative Light Unit	Unități de lumină relative, ce măsoară chemoluminiscența la citometria în flux
RNM		Recuperare Neuromusculară
ROS	Reactive Oxygen Species	Specii reactive de oxygen
RT-PCR	Real time PCR	PCR în timp real
SCP1	Synaptonemal complex protein 1	Proteinei complexului sinaptonemal 1
SCUBA		Spitalului Clinic de Urgență "Bagdasar Arseni"
SDF-1	Stromal cell-derived factor 1	Factorul 1 derivat din celulele stromale
Sema	Semaphorin	Semaforină
SHH- wnt	Sonic hedgehog signaling	Semnalizare pe calea sonic heddgehog
SIRT2		Sirtuina 2
SMAC/ DIABLO	Small mitochondria-derived activator of caspases/ direct inhibitor of apoptosis-binding protein with low pI	Mici activatori ai caspazelor derivați din mitocondrii/ inhibitor direct al proteinei de legătură a apoptozei cu punct izoelectric (pI) scăzut
Smad1	Mothers against decapentaplegic homologue 1	Factor transcriptional
SOD1		Superoxid-dismutaza 1
Sox	Sulfur oxidation	Enzimă oxidantă a sulfului
SCI	Spinal Cord Injury	Traumatism vertebra-medular
TAD	Transactivation domain	Domeniul de transactivare
TAK-1	Transforming growth factor-β-activated kinase 1	Kinaza activată de factorul de creștere transformant-β1
TDO	Tryptophan DiOxygenase	Triptofan dioxigenază

TEHBA	Teaching Emergency Hospital "Bagdasar-Arseni"	Spitalului Clinic de Urgență "Bagdasar Arseni"
TFS		Tampon fosfat salin
TGF-β	Transforming Growth Factor beta	Factorul de creștere transformant-β
TIE1	Tyrosine kinase with immunoglobulin like and EGF like domains 1	Tirozin kinaza cu domenii asemănătoare imunoglobulinei și EGF 1
TIMP1	Tissue inhibitor of metalloproteinase-1	Inhibitorul tisular al metaloproteinazei 1
TLR	Toll-Like Receptor	Receptor Toll-asemănător
TNF-α	Tumor Necrosis Factor α	Factorului de Necroză Tumorală α
TRADD	TNF Receptor-Associated Death Domain	Proteine adaptoare asociate domeniului TNFR-1
TRAF2	TNFR-Associated Factor 2	Factorul 2 asociat TNFR
TRAIL	TNF-Related Apoptosis Inducing Ligand	Ligandul corelat cu apoptoza indusă de TNF-
TrkB	Tyrosine receptor kinase B	Kinaza receptorului tirozinei B
TVM		Traumatism Vertebro-Medular
U87 MG		Linie celulară de glioblastoma
UPR	Unfolded protein response	Răspunsul proteinelor neplicaturate
UV		Ultraviolete
VEGF	Vascular Endothelial Growth Factor	Factorul de creștere endotelial vascular
VEGF-R	VEGF- Receptor	Receptor VEGF
Wnt	Wingless/Integrated	Cale de semnalizare celulară
Wnt/PCP	Wnt/Planar Cell Polarity	
XBP1	X-box binding protein 1	Element component ERSR
XIAP	X-linked inhibitor of apoptosis protein	Inhibitorii proteinelor apoptotice X-linkați

## List of personal published works

1. **Simona Isabelle Stoica**, Ioana Tănase, Vlad Ciobanu, Gelu Onose

*"Initial researches on neuro-functional status and evolution in chronic ethanol consumers with recent traumatic spinal cord injury."*

J Med Life. 2019 Apr-Jun;12(2):97-112. doi: 10.25122/jml-2019-0026. PMID: 31406510; PMCID: PMC6685305. <https://pubmed.ncbi.nlm.nih.gov/31406510>

2. **Simona Isabelle Stoica**, Ioana Tănase, Gelu Onose

*"Influences and consequences resulting in addictions in general and to chronic alcoholism, especially for patients with spinal cord injury"*

Balneo and PRM Research Journal. Vol.12, No.2 June 2021 p: 129–132  
<http://dx.doi.org/10.12680/balneo.2021.432>

3. **Simona Isabelle Stoica**, Coralia Bleotu, Vlad Ciobanu, Anca Mirela Ionescu, Irina Albadi, Gelu Onose, Constantin Munteanu

*"Considerations about Hypoxic Changes in Neuraxis Tissue Injuries and Recovery"*  
Biomedicines. 2022 Feb 18;10(2):481. doi: 10.3390/biomedicines10020481.  
<https://www.mdpi.com/2227-9059/10/2/481>

4. **Simona Isabelle Stoica**, Gelu Onose, Mihail Hoteteu, Constantin Munteanu

*"Effects of ethanol and deferoxamine on rat primary glial cell cultures, in regard with ischemia induced by traumatic spinal cord injury"*

Balneo and PRM Research Journal. 2022, 13(2): 502 <http://bioclima.ro/Balneo502.pd>

5. **Simona Isabelle Stoica**, Ioana Madalina Pitica, Ana Iulia Neagu, Laura Denisa Dragu, Lacramioara-Elena Radu, Lilia Matei, Mihaela Chivu-Economescu, Gabriela Ion, Laura Georgiana Necula, Constantin Munteanu, Aurelian Anghelescu, Gelu Onose, Carmen Cristina Diaconu, Coralia Bleotu

*" The hypoxic stress effect in chronic ethanol exposure of neuronal cells"*

Int. J. Mol. Sci. 2022, 23, x FOR PEER REVIEW

## INTRODUCTION

This PhD thesis comes to the meeting between problems of the modern world (where the active lifestyle predisposes to severe accidents in the conditions of the technological leap) and the consequences of addictive food choices (which have influenced human society since ancient times).[1]. In these conditions, a pathology with increasing frequency is that of vertebral-medullary traumatology, which affects all population categories, with the risk of dysfunction on multiple levels: individual, family, social [2] [3].

We were contradicted by the observations regarding the case report from RNM of the THEBA regarding frequent differences between the objective neuro-dysfunctional state after spinal cord injury (SCI) between patients known to abuse chronic ethanolic and those who did not have such behavioral disorders[4]. Thus, with the approval of the Ethics Committee of the Hospital (no. 35517/25.11.2015), we initiated the clinical study related to this doctoral thesis with the aim of researching the relationship between alcoholism and neuro-dysfunctional and evolutionary recuperative, neuromuscular status after SCI.

The working hypothesis refers to the objective-statistical verification and, as far as possible, through paraclinical investigations including at the intimate tissue and molecular level of the positive (beneficial) influence of chronic alcoholism on the evolution (acute and subacute) of the neuro-dysfunctional clinical status in patients with SCI, where chronic ethanol abuse is known to cause structural and functional damage in nervous tissue.

The general objective of this doctoral study is the analysis and attempt to explain the above-mentioned, somewhat surprising, phenomenon in the context of the verification of the Working Hypothesis.

The specific objectives are:

- the objectification of the statistical significance of the clinical finding from the working hypothesis (through the retrospective analysis of the cases of patients with SCI, hospitalized in the RNM Department of THEBA between 01.01.2005 and 01.06.2022);
- the prospective attempt to explain and identify, through fundamental research approaches, the intimate mechanisms possibly involved in the generation of the (paradoxically) more favorable evolution of myelic tissues under conditions of aggression, in immortal tumor cell cultures;

- the prospective attempt to explain and identify, through fundamental research approaches, the intimate mechanisms possibly involved in the generation of the (paradoxically) more favorable evolution of myelic tissues under conditions of glial cell aggression from primary (rat) cell cultures.

The work is structured in two sections:

- The general part that reviews current scientific data on the researched fields (spinal cord injury, the influences of chronic ethanol impregnation on the human body in general and on the central nervous system - with a main focus on the spinal cord - in particular and a brief systematization of the main mechanisms related to the molecular biology of after SCI injury events, and their interference with neurofunctional clinical evolution, including recovery in the stages from acute to subacute).
- The personal contributions include a clinical-statistical retrospective analysis of the studied problem and a prospective experimental research of the dynamics at an intimate level in neuronal and gliocyte biology, including from the spinal level in experimental conditions with elements prone to the reproduction of conditions similar/partially similar to those in the spinal cord traumatic lesions.

The first step was to document our clinical observations by evaluating the state of knowledge of the issue at the level of the international scientific community [4]; then we carried out an exhaustive retrospective clinical study of patients hospitalized in the THEBA RNM Department with diagnoses of tetraplegia or paraplegia post-SCI (from the archive of the mentioned Clinic in the period 01.01.2005- 01.06.2022) creating a database that includes 1057 such cases , where we performed the statistical analysis of the objective data extracted from the clinical observation sheets of the respective hospitalized persons, including using the clinical-functional evaluation parameters quantified by the specific measurement scales/grids (AIS, Frankel). The retrospective analysis of the data related to the 1057 patients revealed to us that they do not present a normal distribution (taking into account the limiting selective criteria applied), but there are statistically significant correlations between chronic ethanol abuse and the status, as well as the favorable evolution, from the point of view motor vision, as well as the yield of motor and sensory recovery in such spinal cord injury patients.

Later, we carried out a prospective study on neuronal and glial tumor cell lines (in the Department of Molecular and Cellular Pathology of the "Ștefan S. Nicolau" Institute of Virology) on models of cellular suffering (through mechanical trauma and hypoxemic

interventions) in neural cell cultures HTB-11 and U87 MG tumors treated excessively with ethyl alcohol and corresponding in time to alcoholism. We observed morphological differences between neural cells (neurons and gliocytes) grown with and without ethanol treatment, as well as between cells exposed and not exposed to traumatic conditions (through scratching and hypoxia) but without these structural differences jeopardizing cell survival. Then we analyzed (comparing between cells exposed and not exposed to ethanol treatment) the molecular response by studying the chronological post-traumatic cellular events (as known from the literature; initially, after the occurrence of a vertebral-medullary trauma, cellular necrosis processes take place, which - by affecting the membrane excitability - it causes the efflux of potassium in the extracellular environment - with impact depolarization - and the influx of calcium, followed by the excess production of glutamate, which causes a state of local hyperexcitability, which - along with ischemia through bleeding and vaso- spasticity - causes hypoxic changes, which quickly induce metabolic dysfunctions and local oxidative stress, with the activation of the innate immune response at the inflammasome level, with a defensive role, being followed by the initiation of apoptotic mechanisms - involved in both neuroprotective and neurodegenerative processes - as well as influencing molecules of cellular signaling, with the functional reactivation of some embryonic neural pathways). The molecular biology evaluation was carried out in the proteosome conceptual paradigm, by evaluating gene expression (at messenger RNA level) and protein synthesis, specific to the posttraumatic chronological stages described above. Gene expression was assessed by real time polymerase chain reaction (RT PCR) and protein synthesis was quantified by the DotBlot technique, which used specific antibodies. The cell cycle was also evaluated by flow cytometry and the images of the cell cultures were made under an inversion and fluorescence microscope (after special stainings).

Ethanol-impregnated cells (although having a lower resistance to stress) showed regenerative capacity to refill gaps in the scratch test similar to untreated ethanol-treated cells.

Evaluation of the response to hypoxia (targeted by molecular markers: Hypoxia Inducible Factor-1 $\alpha$  – HIF-1 $\alpha$ , Nuclear Factor Kappa-light-chain-enhancer of activated B cells-beta - NF $\kappa$ - $\beta$ , Vascular Endothelial Growth Factor- VEGF, Fms related receptor tyrosine kinase -1 – FLT-1, Transforming Growth Factor beta - TGF- $\beta$ , Protocadherine12 - PCDH12, Hemoxigenase1 - HMOX1, Hemoxigenase2 - HMOX2, Carbonic Anhydrase 9 - CA9, Hypoxia Inducible Factor-2 $\alpha$  - HIF-2 $\alpha$ , Indoleamine 2,3-DiOxygenase - IDO) of

neuroblastoma cells exposed to chronic ethanol treatment showed that the following effects of ethyl alcohol occur: reduction of neurodegeneration and neurotoxicity functional improvement of the response/metabolic-functional resilience of neurons, therefore neuroprotective, anti-inflammatory, anti-apoptotic, angiogenesis, of synaptic remodeling/neuroplasticity.

Cell stress study (through the molecules: interleukin-1 $\beta$  - IL-1 $\beta$ , TNF- $\alpha$  - Tumor Necrosis Factor, interleukin-6 -IL-6, interleukin-8 -IL-8, interleukin-4 -IL-4, Catalase, Claspin , Clusterin, Hat Shock Protein27- HSP27, Hat Shock Protein60 - HSP60, Hat Shock Protein70 - HSP70) showed that chronic exposure to ethanol induces (on neural cells treated with hypoxemic agents) the following effects: antioxidant, adaptive regulation of energy metabolism, neuroprotective, anti-inflammatory, antiapoptotic, anti-scarring, nerve regeneration inhibitor as a defensive response, to restore nerve endings, to inhibit axonal dieback phenomena.

Cell stress study (through the molecules: interleukin-1 $\beta$  - IL-1 $\beta$ , TNF- $\alpha$  - Tumor Necrosis Factor, interleukin-6 -IL-6, interleukin-8 -IL-8, interleukin-4 -IL-4, Catalase, Claspin, Clusterin, Hat Shock Protein27- HSP27, Hat Shock Protein60 - HSP60, Hat Shock Protein70 - HSP70) showed that chronic exposure to ethanol induces (on neural cells treated with hypoxemic agents) the following effects: antioxidant, adaptive regulation of energy metabolism, neuroprotective, anti-inflammatory, antiapoptotic, anti-scarring, nerve regeneration inhibitor as a defensive response, to restore nerve endings, to inhibit axonal dieback phenomena.

Chronic ethanol treatment of immortal neuronal cultures (of neuroblastoma), exposed to hypoxemic experimental conditions, produced the following consequences at the level of cellular carbohydrate metabolism (through the molecules: Adenylkinase3 - AK3, Fetuin-B - FETU-B, Phosphoglycerate Kinase - PGK1, Proprotein Convertase Subtilisin/Kexin type 1 - PCSK1, PhosphoFructoKinase, Liver Type 1 - PFKL1, High Mobility Group Box 1 - HMGB1, Glucose transporter 1 - GLUT1, Tyrosine kinase with immunoglobulin like and EGF like domains 1- TIE1, Lactate DeHydrogenase B - LDH B ): energy depletion and increase in lactic acid level, but also stimulation of the proteolytic activity of insulin, improvement of the yield of glucose use, favoring its intracellular penetration, and (including) vascular remodeling.

Apoptosis study (through the molecules: Caspases -3, -7, -8, -9, -10, HTRA2/Omi, Livin, p21, p27/Kip1, Phospho-p53 (S15), Phospho-p53 (S46), Phospho-p53 (S392), Phospho-Rad17 (S635), Small mitochondria-derived activator of caspases/ direct inhibitor of apoptosis-binding protein with low pI - SMAC/Diablo, Survivin, Tumor Necrosis Factor Receptor - TNF R, X-linked inhibitor of apoptosis protein – XIAP, Bad, Bax, Bcl-2, Bcl-x, Pro-Caspase-3, Cleaved Caspase-3, Catalase, Inhibitor of apoptosis proteins-1 - cIAP-1, Inhibitor of apoptosis proteins 2 - cIAP-2, Claspin, Clusterin, Cytochrome C, TNF-Related Apoptosis Inducing Ligand1 -1- TRAIL R1, TNF-Related Apoptosis Inducing Ligand2 - TRAIL R2, Fas-Associated Death Domain - FADD, Fas) showed that chronic exposure to ethanol causes antiapoptotic effects (in the extrinsic and intrinsic way), including in conditions of hypoxemic interventions.

Evaluation of intercellular signaling molecules (Laminin4 - LAMA4, Chitinase 3-like protein 1 - CHI3L1, Lipocalina2 - LCN2, Tissue inhibitor of metalloproteinase-1 - TIMP1, Semaphorin3 - SEMA3) and embryonic communication pathways (Sonic hedgehog signaling - SHH, Prominin -1 - CD133, Neurogenic locus notch homolog protein1 - Notch1, Hairy Ears, Y-linked – Hey, Bone morphogenetic proteins 4 - BMP4, Hairy/enhancer of split – Hes, Wingless/Integrated – WNT, Caderina 2 - CDH2, Vimentina – Vim, Nestina – Nestin, Neuronal differentiation 1 - NeuroD1, Glial fibrillary acidic protein - GFAP, Oligodendrocyte transcription factor 2 - Olig2) in neurons (neuroblastoma type) chronically treated with ethyl alcohol revealed the following consequences: reduction of neuraxial scars; stimulation of neurite development, vascular remodeling, neurogenesis, synaptogenesis, neuroregeneration; increasing the ability to differentiate into neurons or glial cells.

As the study on tumor cells presents the great disadvantage of significant genetic and metabolic differences, compared to non-tumor myeloid cells, we carried out (in the RNM SCUBA Research Core) another prospective study on the relationship between ethanol exposure and (spinal-)traumatic heart suffering in cell cultures newborn rat primaries cultured in ethanol exposure (trying to reproduce, as far as possible, the temporal correspondence with chronic alcoholism in humans). The analysis was carried out by evaluating the protein synthesis (using the ELISA test) at the level of the inflammasome (with the evaluation of IL-6) and the apoptosis process (by determining TNF- $\alpha$ ) in the cells of primary cerebro-spinal glial cultures from the rat. We observed that, in primary cultures with such glial cells, the stress induced by ethanol treatment, supplemented by hypoxia,

accentuates inflammatory intercellular communication, but without leading to marked levels of programmed cell death.

The last aspect of our research concerned the prospective analysis of phenomena at the human biological level, which is why we analyzed the distribution of cytokines (inflammatory and proapoptotic) in the cerebrospinal fluid collected from several patients with damaged dura mater following a SCI (possible collection for bioethical reasons only in such situations, which explains the small number of these cases).

Experimental evaluation of CSF was performed by electrophoresis and ELISA method for TNF- $\alpha$  and IL-6. The analysis of CSF in patients chronically consuming excess ethanol revealed the presence of pro-inflammatory and pro-apoptotic tendencies, possibly including as a mechanism of isolation and protection of the injured medullary area, of course the number of cases studied being insufficient for processing and statistical analysis.

Overall, this doctoral thesis addresses a subject we consider particularly interesting and encountered only tangentially - concretely we did not find it studied from the perspective of our working hypothesis - in the literature, which, especially under the conditions in which, unfortunately, in currently there is still no form of therapeutic intervention able to effectively heal spinal cord injuries, including traumatic ones (their neurodysfunctional consequences are not infrequently devastating due to their huge disabling potential, often for life), opens a way to explore a field which, under the conditions of additional deepening, could lead to the identification of contributory therapeutic-restorative solutions.

## **I. GENERAL PART**

### **1. General data on spinal cord injury**

SCI "represents the cause of a plegic deficit, with temporary or permanent loss of nervous control over a somatic and vegetative territory", caused by damage to the spine and spinal cord (and spinal nerves), after: acts of violence, road accidents, falls, sports accidents[4] [5] [6]. The incidence of TVM is 10.4–83 cases/million patients/year (ie approximately 768 473 annual cases), affecting mostly patients in the age group of 30 years [7], being a complex challenge (through its individual, family consequences and social) [8] [9]. Mortality after SCI is directly proportional to the height of the lesional level (spinal cord), the advanced age of the victims and the force of the traumatic impact [9].

In the body, SCI produces various changes: represented by primary injuries (the consequences of the traumatic impact force on the spine: flexion, extension, rotation, compression; all affecting the spinal cord, causing medullary contusion, with transient or permanent neurological dysfunctions, depending on the severity insult) and secondary (in which the nerve suffering from the initially injured area spreads around, especially through the ischemic phenomena produced by vascular injuries, capillary thrombosis, vasospasm and edema; with the appearance of a significant local metabolic and energetic imbalance) [8] [9].

The etiological treatment of traumatic vertebral-medullary disorders is still in the research stage, without known therapeutic molecules capable of inducing the disappearance of motor deficits and sensitivity disorders following SCI.

Recovery after vertebral-medullary traumas is an arduous process (based on the spinal resources of the reorganization of the nervous structures that have survived post-traumatic), of long duration most of the time; being the result of the work of the specialized team (consisting of: recovery doctor, physiotherapist, clinical psychologist, medical assistant, nurse, stretcher bearer, occupational therapist, joined by the patient's family members) by promoting the patient's survival, with the adoption of a lifestyle adapted to the new conditions [11] [12].

## **2. Chronic alcoholism and its physiopathological consequences, respectively complex, in spinal cord injury**

Alcohol (ethyl) is perhaps the cheapest and most used antidepressant by many people over time. The consequences of this fact are multiple, taking on the dimensions of a societal problem. Our brains have reward and tolerance pathways that quickly turn occasional ethanol consumption into a vicious lifestyle. Ethyl alcohol affects the development of the human body in all its stages, starting from intrauterine life, influencing individuals of all age categories, sex, lifestyle or other comorbidities.

We define chronic alcoholism as a long exposure (greater than 7 days/two weeks) to high doses of ethanol: more than 8 standard daily ethanol units (standard drink per day = 10 mg ethanol)[26] [27].

From a molecular point of view, the structures of ethanol and polyethylene glycol (a molecule researched in the treatment of SCI) have many physicochemical properties in common, as they are part of homologous series of the class of alcohols.

Tabel.1. 1. Physico-chemical and structural characteristics of ethanol and PEG (taken from own article [3]) [42,43]

<b>ALCOHOLS/ POLYOLS</b>	<b>ETHANOL (ethyl alcohol, metilcarbinol, ethyl hydroxyl, ethane monoxide, etiolte, hydroxyethane)</b>	<b>POLYETHYLEN GLYCOL (oxid de polyethylen oxide, polyoxyetilene)</b>
Method of production	Fermentation of cereallo/ fruits/ some plants	Industrial biosynthesis procedures
Physical propertes	Liquid, volatile, flammable, characteristic smell	Molecular weight variables: liquid/ solid
Structure	CH <sub>3</sub> -CH <sub>2</sub> -OH	H-(O-CH <sub>2</sub> -CH <sub>2</sub> ) <sub>n</sub> -OH
Molecular weight	46,076844 g/mol	18.02 + 44.05 ng/mol (300- 10000000 g/ mol)
Density	0.7894 g/cm (at 25 Celsius degrees)	Variable
Boiling point	78,37 Celsius degrees	182 – 287 Celsius degrees
Freezing point	-114 ,4 Celsius degrees	Variable
Chemical properties	Water-soluble alcohol	Water-soluble alcohol, methanol, ethanol, acetonitrite, benzene, dichlormetane, acetone. Insoluble in diethyleter, hexane
Biological Effects	It crosses the blood-brain barriere, euphoric, sedative, anxiolytic/ antidepressant, antiseptic, solvent, antitussive, antidote, fuel (7 kcal/gram)	Osmotic laxative, coating film for drug molecules, vector in genetic therapy

Tabel.1.2. Pharmacokinetics of ethanol and PEG (taken from own article [3]) [43–47]

<b>ALCOHOLS/ POLYOLS</b>	<b>ETHANOL</b>	<b>POLYETHYLEN GLYCOL (PEG 400/2000 / 5000)</b>

Absorbption	Stomach, small intestine Peack of blood concentration in 30- 60 min	Intravenous use
Distribution	Anywhere in the body Rate of extraction = 0.2 There are no plasma transport proteins Target: liver, nevrax, heart	0.07% per gram of neural tissue
Elimination	Limited to the maxiumm 8.5 g/h/70 kg Liver metabolism: *alocoholdehidrogenase, *aldehyddehidrogenase Expiratory (0.7%) - 0.16 L/h Urinary (0.3%) - 0.06 L/h Sweting (0.1%) - 0.02 L/h	PEG 2000- 1.4 ml/ min/ kg PEG 5000- 0.4 ml/min/kg

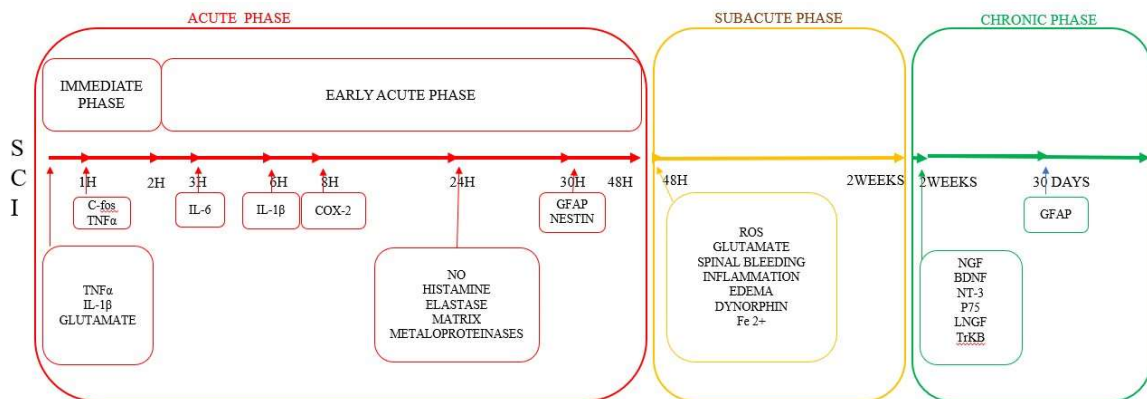
Tabel. 1.3. Pharmacodynamics of ethanol and PEG (taken from own article [2]) [43,47]

<b>ALCOHOLS/ POLYOLS</b>	<b>ETHANOL</b>	<b>POLYETHYLEN GLYCOL</b>
Mechanism of action	NMDA receptor antagonist (Decrease of neurotoxicity due to NMDA receptor antagonist) GABA- A receptor agonist 5HT3 agonist Stimulate opiate receptors and production of dopamine	Decreases intracelluarrelease of oxygene free radicals; stabilizes cell membranes against lipid peroxidation; limits injury secondary membrane lesions
The acute phase is subdivided into an immediate phase (in the first 0-2 hours) and an early acute phase (at 2-48 hours posttraumatic).	The acute phase is subdivided into an immediate phase (in the first 0-2 hours) and an early acute phase (at 2-48 hours posttraumatic).	The acute phase is subdivided into an immediate phase (in the first 0-2 hours) and an early acute phase (at 2-48 hours posttraumatic).

### 3. Molecular biology mechanisms in spinal cord injury and data on current therapeutic possibilities

Spinal cord injury is defined as an acute injury event, occurring at the level of the spinal cord and its adjacent structures. Post-traumatic changes are manifested from the initial stage, which extends between 2-48 hours post-injury [48]. The following stages after SCI are described: acute phase (0-48 hours after SCI), subacute (between 2 days and 2 weeks post-traumatic) and chronic (following the subacute phase) [49]. In the immediate acute phase, axonal ruptures, hemorrhages in the gray matter, local ischemia, neuronal death occur. Now the microglia are activated, with the release of TNF- $\alpha$ , IL-1 $\beta$  (even from the first minutes after the injury) and with the production of glutamate in cytotoxic amounts [50].

The structural and functional changes induced by SCI originate in nerve cell dysfunctions at different levels, starting with the process of cell division (in the case of glocytes and neural stem cells).



*Fig. 1.1. Evolutionary phases after SCI and molecular changes produced in nervous tissue. The following stages are distinguished according to SCIS: the acute phase (in the first 48 hours post-traumatic) consists of the immediate phases (which happens in the first 2 hours after SCI) and the early acute phase (between 2 hours and 48 hours post-traumatic), the subacute phase (with a duration of 2-14 days after SCI) and the chronic phase (which begins after the first 2 post-traumatic weeks). At the time of TVM production, the following are released (in the nerve tissue): TNF $\alpha$ , IL-1 $\beta$ , glutamate; one hour post-traumatic, the following are released: c-fos, TNF $\alpha$ ; 3 hours after TVM: IL-6 is released; at 6 hours IL-1 $\beta$  is released; at 8 hours: COX-2 is produced; and at 24 hours: nitric oxide, histamine, elastase are synthesized, matrix metalloproteinases; while 30 hours post-traumatic: GFAP and nestin are produced; 48 hours after TVM are produced (on the background of spinal bleeding and local edema): reactive oxygen species, dynorphin and ferric ions; after 28 days posttraumatic are produced: NGF, BDNF, NT-3, LING, TrKB, and 30 days after the SCI, GFAP is synthesized.*

Hypoxia is defined as the decrease or deprivation of oxygen at the level of organs, tissues, cells by decreasing the oxygen supply (due to damage to the vascular network, anemia) or by increasing oxygen consumption (as in the sudden increase in the rate of cell proliferation)[58].

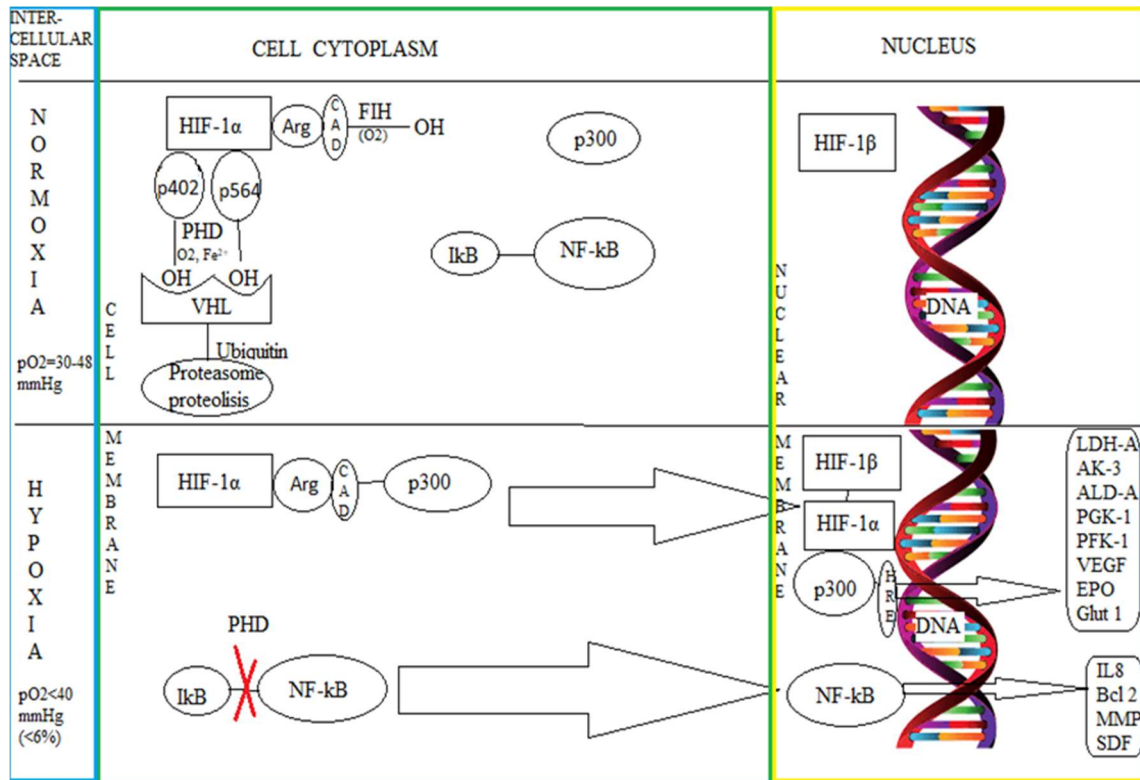


Fig. 4.1. Hypoxic cellular mechanisms with the influence of oxygen partial pressure,  $pO_2$ , on deoxyribonucleic acid, DNA, HIF, with participation, FIH; ARG; CAD; protein 300, p300, VHL and PHD. We observe the activation (on the atypical pathway) of NF- $\kappa$ B (with the involvement of I- $\kappa$ b), of HRE with the stimulation of gene expression: LDH-A; AK-3; ALD-A; PGK-1; PFK-L; VEGF; EPO; Glut-1; IL-8; Bcl-2; MMP; SDF (drawing taken from own article[209])

The inflammasome is a multiprotein cytosolic complex that functions as an intracellular receptor for cellular and environmental stress [98] mediating the innate immune response, which can cause tissue damage when overactivated [99] [100].

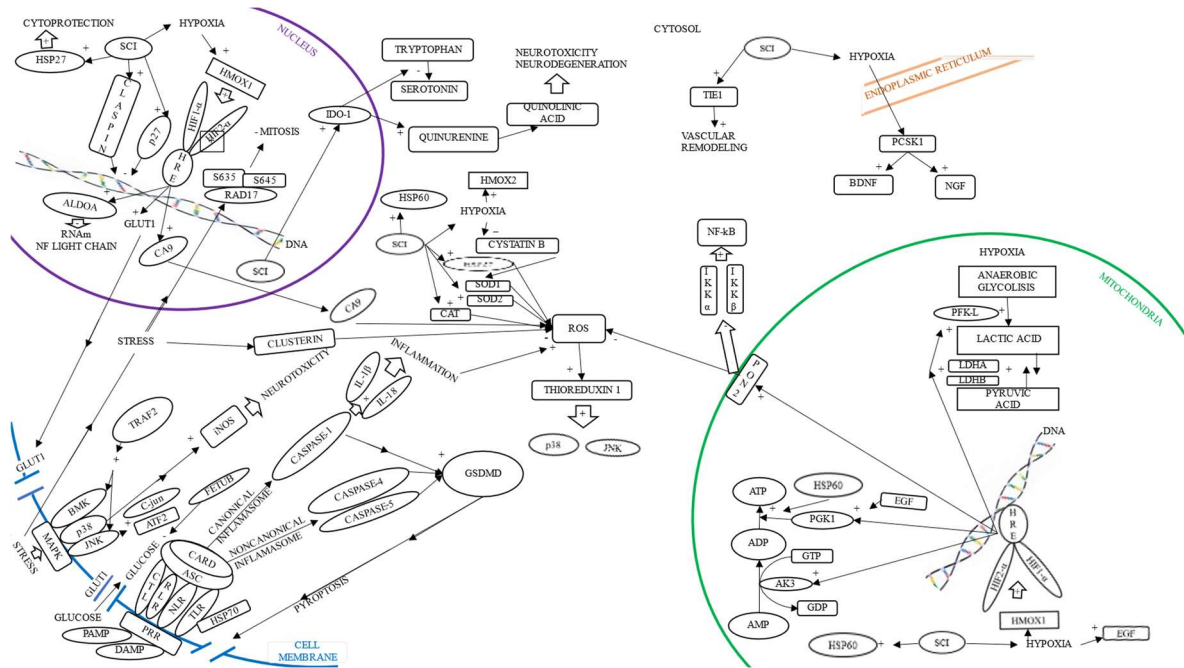


Fig. 1.2. Baseline metabolic and inflammatory changes after SCI. Events in SCI lead to the appearance of DAMPs and PAMPs (in the intercellular space), which interact with PRRs, then with CTLs, RLRs, NLRs and TLRs (whose activity is also controlled by HSP70), which causes the activation of ASCs (and the CARD of its interior), followed by caspase interaction: with caspase-1 (which leads to inflammation by stimulating IL-1 $\beta$  and IL-18, via the canonical inflammasome activation pathway) or with caspases -4 and -5 (which stimulate GSDM and pyroptosis on the non-canonical inflammasome activation pathway). The stress produced by SCI can also activate membrane MAPK, which then initiates the p38 (with stimulation of iNOS leading to neurotoxicity) and BMK (which is stimulated by TRAF2) and JNK (activated by TRAF2, which stimulates cjun and ATF2) pathways. The stress produced after SCI can also act at the nuclear level, via RAD17 (which is phosphorylated at S635 and S645, inhibiting mitotic activity). Also at the nuclear level, SCI can lead to the transcriptional activation of IDO-1 which can inhibit the synthesis of serotonin from tryptophan or stimulate the quinic acid pathway (with the risk of producing neurotoxicity and neurodegeneration). But stress can also activate cytoplasmic clusterin which can inhibit ROS (with an antioxidant role). Activated ROS can stimulate thioredoxin1, which can stimulate JNK and p38 proteins in the MAPK pathway. SCI at the nuclear level can also stimulate the synthesis of HSP27 (with a cytoprotective role), claspin and p27 (which inhibits DNA activation). And hypoxia following SCI influences DNA (nuclear and mitochondrial) by stimulating HMOX1, which causes increased levels of HIF-1 $\alpha$  and HIF-2 $\alpha$ , which then activates HRE, which then causes synthesis of ALDOA (with stimulation of neurofilament light chain mRNA), of GLUT1 ( which favors the entry of glucose inside the cell, and FETUB produces resistance to the action of glucose), of CA9. At the mitochondrial level, SCI stimulates HSP60, and the subsequent hypoxia leads to the stimulation of HRE that controls cellular energy metabolism by being involved in ADP phosphorylation (stimulating AK3), in ADP phosphorylation (stimulating PGK1), in lactic acid production (through PFK-L synthesis), in bilateral lactate-pyruvate conversion (through the synthesis of LDHA and LDHB). Mitochondrial hypoxia also stimulates EGF with possible consequences in vascular remodeling. And HRE also stimulates PON2, resulting in NF- $\kappa$ B activation (by inhibiting IKK $\alpha$  and IKK $\beta$ ). On the other hand, SCI can inhibit ROS by stimulating SOD1 and SOD2, CAT. At the cytosolic level, SCI can stimulate TIE1 (involved in stimulating vascular remodeling); and at the level of the endoplasmic reticulum, it increases the production of BDNF and NGF (via PCSK1).

Cell death, according to the classification of the Nomenclature Committee on Cell Death (NCCD) from 2018, can be of two types: accidental (accidental cell death - ACD) and

programmed/regulated (regulated cell death - RCD)[120][9]. ACD is a form of instantaneous cell death produced by the disintegration of the cell membrane through the action of aggressive factors (physical, chemical or mechanical), and RCD is the consequence of the activation of one or more cellular signaling pathways (which can be influenced pharmacologically) [9][120].

After SCI, cell death occurs through both pathways: ACD and RCD [120]. Neural cell death is achieved by: apoptosis, necrosis, necroptosis, ferroptosis, pyroptosis, autophagy, paratantosis, oncosis [121][122] [123] [55] [9][120][124].

Apoptosis is a highly regulated, non-inflammatory process of programmed cell death essential for permanent renewal (at the cellular and whole-organism level)[125] [124] [126].

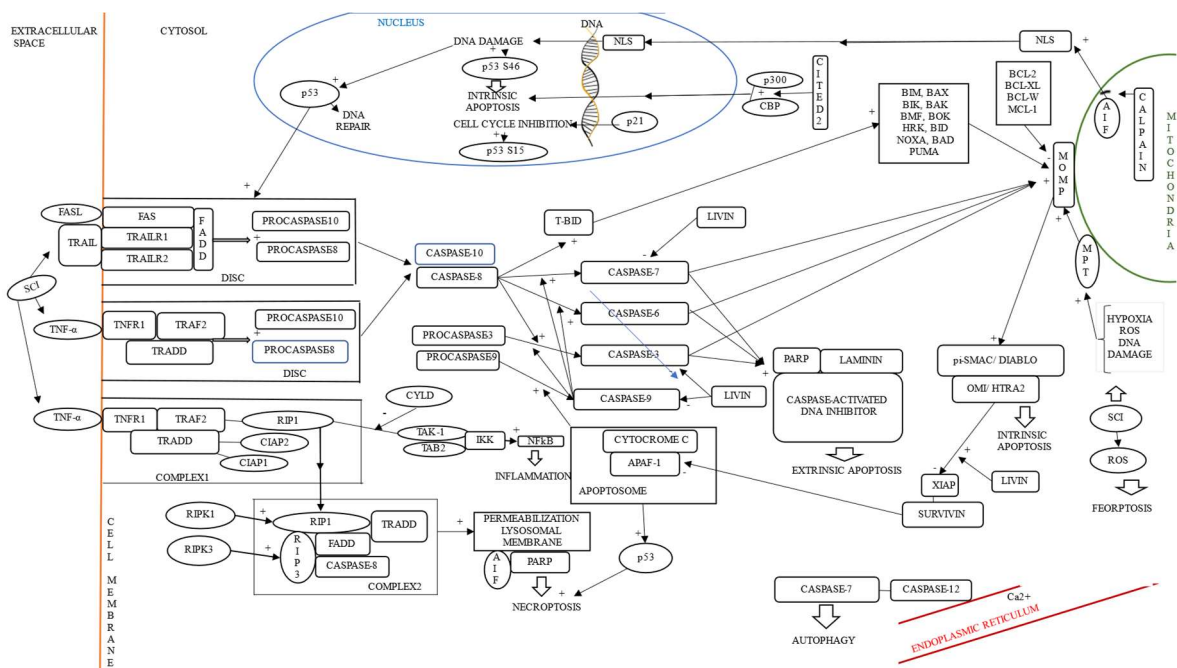


Fig. 1.3. Pathways of cell death in SCI. SCI activates apoptosis through the extrinsic and intrinsic pathway. Thus, FASL and TRAIL act on the DISC (consisting of FAS, TRAILR1 and TRAILR2 that interact with FADD, activating procaspase-8); similar to TNF $\alpha$  acting on the DISC (consisting of TNFR1, TNFR2 interacting with TRADD, activating procaspase-8). DISC then activates caspase-8 which stimulates effector caspases (-3, -6, -7), which act on PARP, laminin, and caspase-activated DNA inhibitor, triggering apoptosis via the extrinsic pathway. Caspase-8 (via T-BID) stimulates proapoptotic genes (BIM, BAX, BIK, BAK, BMF, BOK, HRK, BID, NOXA, BAD, PUMA) which, together with MPT (whose expression is stimulated by hypoxia, ROS and DNA damage) activates MOMP which (mediated by pi-SMAC/ DIABLO and OMI/ HTRA2) triggers the intrinsic pathway of apoptosis and inhibits XIAP which (together with survivin) inhibits APAF-1 (which makes up the apoptosome together with cytochrome C). SCI, via ROS, also stimulates ferroptosis. The apoptosome activates necroptosis (mediated by p53) and stimulates the transformation of procaspase-9 to caspase-9 which stimulates the effector caspases of the extrinsic apoptotic pathway (-3, -6, -7) that also activate MOMP (being also involved in the intrinsic apoptotic pathway). The balance in the intrinsic apoptotic pathway is maintained with the help of antiapoptotic genes (BCL-2, BCL-XL, BCL-W, MCL-1) and livin protein (which inhibits caspases -3, -7, -9 and stimulates XIAP). TNF $\alpha$

also activates inflammation by acting on inflammasome complex 1 (consisting of TNFR1, TNFR2, TRADD, RIP1, CIAP1, CIAP2) which (together with RIPK1 and RIPK3) acts on inflammasome complex 2 (consisting of RIP1, RIP3, FADD, TRADD and caspase-8). Inflammasome complex1, via TAK-1, TAB2, IKK (inhibited by CYLD) stimulates NF- $\kappa$ B triggering inflammation. Inflammasome complex2, by permeabilizing the lysosomal membrane and AIF, PARP triggers necroptosis. Mitochondrial AIF causes activation of cytoplasmic NLS (which causes nuclear DNA damage); and the action of AIF is inhibited by calpain. SCI can produce nuclear DNA damage that stimulates p53 expression (with activation of DNA repair but also with activation of the extrinsic apoptotic pathway) and phosphorylation of p53 at serine 46 (with activation of apoptosis via the intrinsic pathway). Phosphorylation of p53 at serine 15 can be caused by cell cycle inhibition under the action of p21. The intrinsic apoptotic pathway is also activated by the action at the DNA level of the p300-CBP complex (stimulated by cited-2). The endoplasmic reticulum can also be involved in apoptosis, which can attach caspase-12 and caspase-7, triggering the autophagy process.

Looking at posttraumatic neuraxial structural changes, at a microscopic level, the central nervous system is a complex structure made up of cells that are embedded in a soft network of polysaccharides (consisting of proteins and glycosaminoglycans, the best known being: laminin, fibronectin and collagen IV)[138].

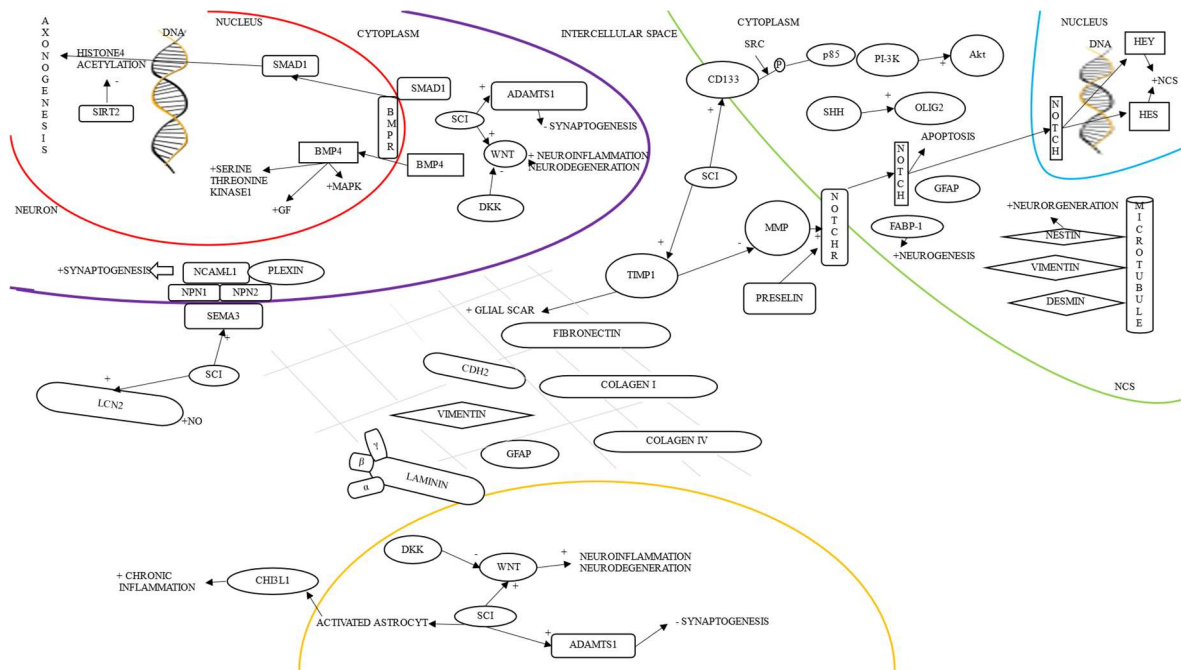


Fig. 1.4. Intercellular signaling after SCI. SCI activates the astrocytic cell which: synthesizes CHI3L1 (promoting chronic inflammation in the intercellular space), stimulates ADAMST1 (with inhibition of synaptogenesis), activates the WNT pathway (leading to neuroinflammation and neurodegeneration, being inhibited under the action of DKK). At the neuronal level, SCI activates the WNT pathway, stimulates ADAMST1, reactivates the SEMA3 pathway (which binds to NPN1, NPN2, NCAM-1 and PLEXIN receptors, stimulating synaptogenesis), acts on BMP receptors via BMP4 (with stimulation of MAPK, increase of serine threonine kinase1), of SMAD1 (with histone 4 acetylation that determines axonogenesis, a process inhibited by SIRT2). SCI also influences neural stem cells by activating: membrane CD133 (which, together with SRC, phosphorylates p53 and PI-3K which activates Akt), TIMP1 (which inhibits MMP, which together with preselin acts on NOTCHR activating the NOTCH pathway which causes apoptosis or activation of Hes and Hey genes causing further activation of NCS), microtubule interaction with vimentin, desmin and nestin (promoting neurodegeneration), FABP-1 (stimulating neurogenesis), SHH (activating OLIG2) and

*GFAP. At the level of the intercellular space, SCI stimulates the formation of glial scars (under the action of TIMP1) which are composed of: laminin (with  $\alpha$ ,  $\beta$ ,  $\gamma$  chains), GFAP, collagen I and IV, fibronectin, CDH2 and vimentin; scar that limits post-traumatic nerve regeneration, but also diminishes the local inflammatory process.*

Different medicinal substances and therapeutic methods are being studied with a potentially beneficial role in ameliorating the deficits that occur after the occurrence of SCI. Substances such as: non-steroidal anti-inflammatory drugs, group B vitamins, lithium, antioxidant molecules (vitamins A, E, C, coenzyme Q10), iron chelating molecules (deferoxamine), statins, inhibitors are considered to favor the neurorecovery process (including after brain trauma) of phosphodiesterase (methylxanthines - theophylline), acrolein antagonists (acetylcysteine acid), neurite growth promoters (Keltican), neurotrophic molecules (Cerebrolisyn, Actovegin), erythropietin, neuronal antiexcitatory molecules (Memantine, Riluzole), calcium channel blockers [20] [6]. At the same time, research is being done on a class of therapeutic molecules with a positive effect in the recovery of S(C)-I: that of polyethylene glycol (PEG). PEG applied to the site of spinal injury (immediately and 8 hours post-injury) in severe S(C)I – produced by spinal cord compression [195] – in adult guinea pigs determined very good neuro-functional consequences (in 23 out of 25 treated animals intralesional, intravenous and subcutaneous with posttraumatic PEG), by a protective effect against damage to axonal membranes in spinal neurons of experimental animals[196–198].

## **II. PERSONAL CONTRIBUTIONS**

### **4. Working hypothesis and general objectives**

The clinical experience acquired after long periods of work with vertebral-medullary traumatized patients led us to note clinical-evolutionary differences after SCI (between acquaintances with chronic ethanol abuse and those with a normal diet, which led us to initiate a research detailed description of the issue (in a doctoral study).

The working hypothesis of this doctoral thesis refers to the positive (beneficial) influence of chronic alcoholism on the evolution (acute and subacute) of the clinical condition in patients with SCI, because chronic ethanol abuse causes structural and functional impairments in the nervous tissue.

The general objective of this doctoral study is to analyze and demonstrate the validity of the Working Hypotheses.

The specific objectives are:

- demonstration of the statistical significance of the working hypothesis (through the retrospective analysis of the cases of patients with SCI);
- prospective demonstration of the working hypothesis at the level of neuronal cells from (immortal) tumor cell cultures;
- prospective demonstration of the working hypothesis by studying glial cells from primary (rat) cell cultures.

## **5. General research methodology**

Initially, we performed a retrospective study of the data from the archive of the RNMM SCUBA Clinic and processed the data through statistical analysis, using the IBM SPSS Statistics v22 program. Some graphics were made with the help of the MS Office 365 Excel program.

The evaluation of the working hypothesis was performed prospectively starting from experimental models that intended to reproduce the neural distress caused by SCI. Thus, considering the post-traumatic damage (in various evolutionary moments) of the vasculature of the spinal cord structures (with the occurrence of tissue ischemic phenomena), we considered the role of cellular hypoxia in the neural lesion evolution to be very important. Hypoxia influences both neural cell metabolism and structure, as well as neuraxial morphophysiology. Therefore, we chose to reproduce (for prospective studies on cell cultures) posttraumatic suffering by inducing hypoxia (through treatments with specific molecules: deferoxamine and cobalt chloride). The prospective study was performed on neonatal rat primary neural tumor and neuraxial cell cultures. Molecular biology determinations and CSF samples of operated SCI patients were performed.

## **6. Study 1: Retrospective data analysis**

We researched the archive of the Neuro-Muscular Recovery Clinic of the "Bagdasar Arseni" Hospital between 01.01.2005- 01.06.2022 and obtained data for a number of 1057, admitted and treated on the ward for acute and subacute status after SCI.

The study showed (with statistical significance) that the motor status after SCI is superior for chronic ethanol consuming patients, as is the performance of sensory-motor recovery processes.

Table 6.48. The result of statistical comparisons of independent numerical data AIS score according to chronic ethanol consumption

		Consum etanolic cronic	N	Media	Diferența mediilor	Valoare p
Scor AIS motor	la internare	Nu	816	41.1691	8.3869	<0.0001
		Da	241	49.556		
	la externare	Nu	816	43.0294	9.3772	<0.0001
		Da	241	52.4066		
	Delta	Nu	780	4.364	3.3386	<0.0001
		Da	232	7.7026		
Scor AIS senzitiv	la internare	Nu	816	119.7868	4.1966	0.738
		Da	241	123.9834		
	la externare	Nu	816	125.6814	6.3062	0.104
		Da	241	131.9876		
	Delta	Nu	814	7.8958	1.9036	<0.0001
		Da	241	9.7994		

## 7. Study 2: Experimental evaluation of neurons from tumor cell cultures

The desire to clarify the relationship of the nervous system with ethyl alcohol (especially in traumatic conditions), also directed us to the study of the behavior of neuronal cells, which we exposed for a long time (chronically) to ethanol. Experimentally, we created a model of neural suffering (also present in traumatic conditions due to the ischemic phenomena that occur) by inducing cellular hypoxia.

We subsequently evaluated comparatively (between neuronal cells exposed and not exposed to ethanol) cell behavior and characteristic molecular biology changes. Research was conducted on the ethanol-exposed neuroblastoma cell line HTB-11 (hereafter referred to as SK-N-SH). The viability study showed that a concentration of 50mM ethanol does not significantly affect cell viability (98.7% cells being viable) and for this reason it was used as a chronic treatment with ethanol (and carrying out treatments for 2 weeks and for more than 9 weeks of cell cultures). The hypoxemia treatment used hypoxia inducers: cobalt chloride (CoCl<sub>2</sub>) and deferoxamine (DFX) in concentrations of 50 μM and 100 μM. The scratch test evaluation is shown in Fig. 7.2.

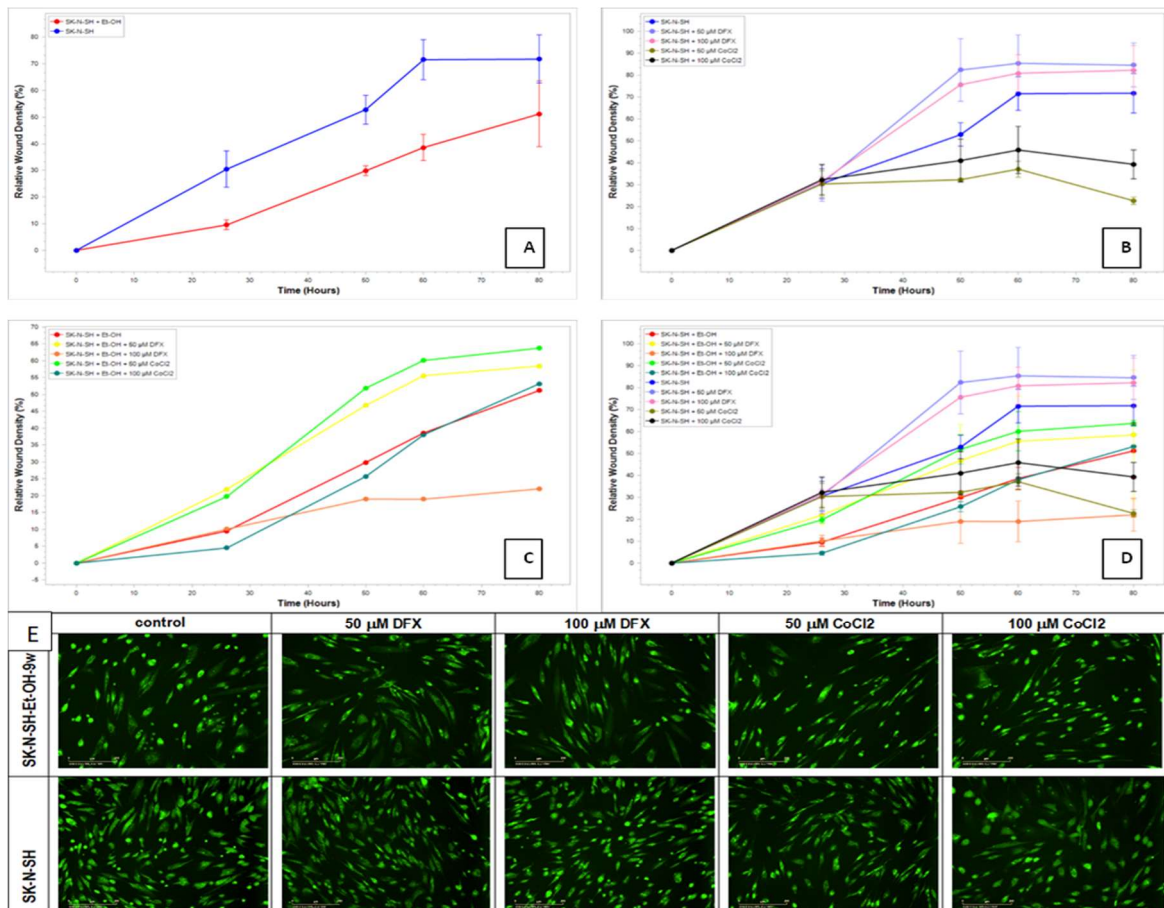


Fig. 7.2. Proliferation and migration of SK-N-SH cells: A. treated (9w) and untreated with ethanol; B. treated (for 24 hours) with DFX and CoCl<sub>2</sub> 50/100  $\mu$ M after scratching; C. exposed 9w to ethanol, then treated (for 24 hours) with DFX and CoCl<sub>2</sub> 50/100 $\mu$ M; D. treated 9w with ethanol, scratched, then treated (24h) with DFX and CoCl<sub>2</sub>. E. Acridine Orange/Propidium Iodide Stain

The production of reactive oxygen species (ROS) was also analyzed, which at low concentrations of hypoxic inducers (50 $\mu$ M CoCl<sub>2</sub> and DFX) had minimal values for cells maintained in ethanol for more than 9 weeks.

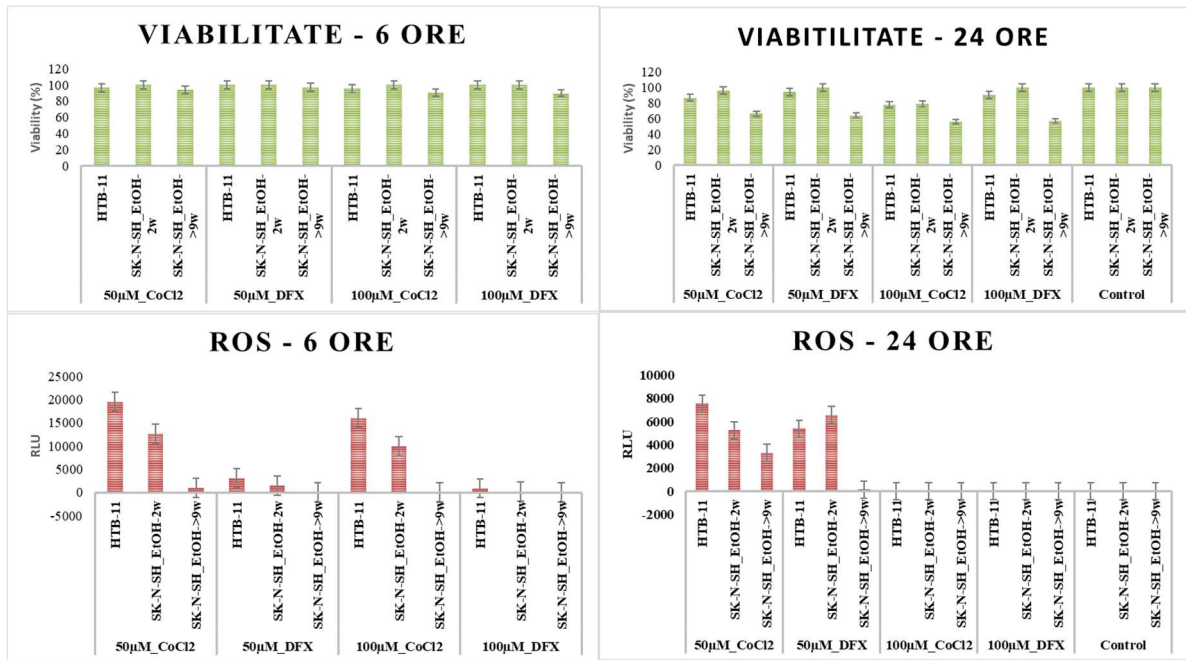


Fig. 7.6. (a, b, c, d) Quantification of cell viability and ROS at 6 and 24 hours

Gene expression of proteins involved in the hypoxic response is shown in Fig. 7.7.

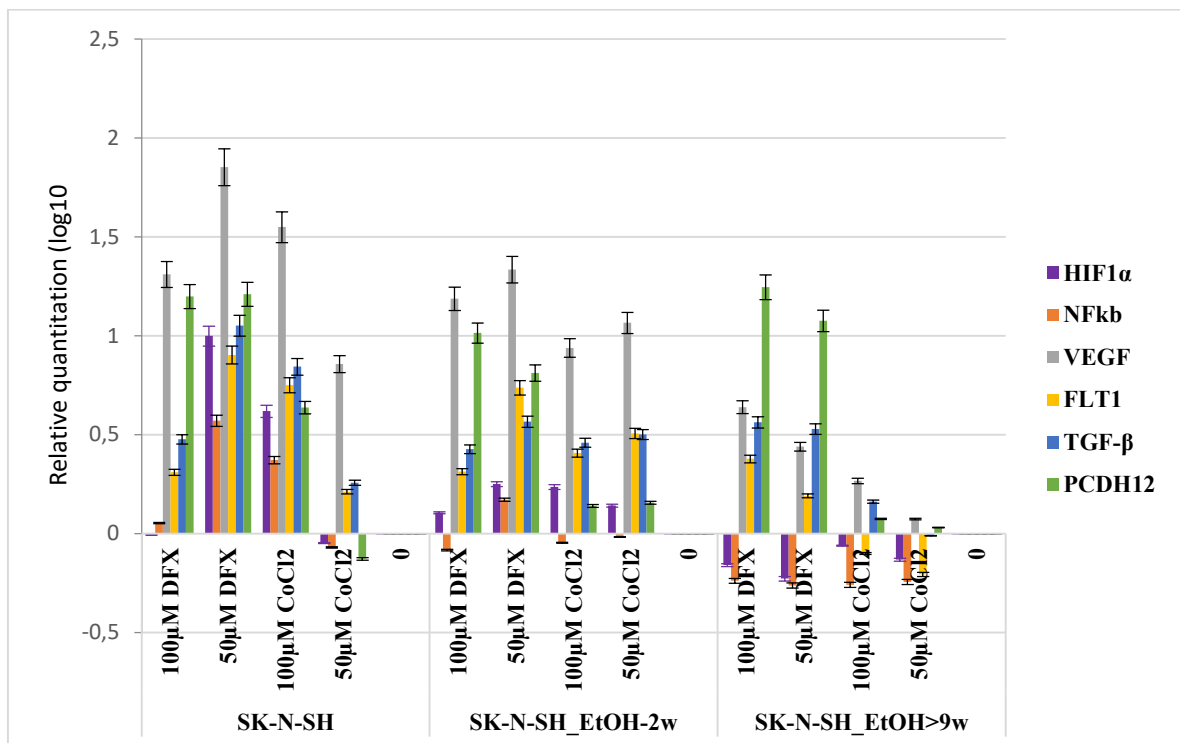


Fig. 7.7. RT PCR Hypoxia

Gene expression of cellular stress proteins is shown in Fig. 7.10.

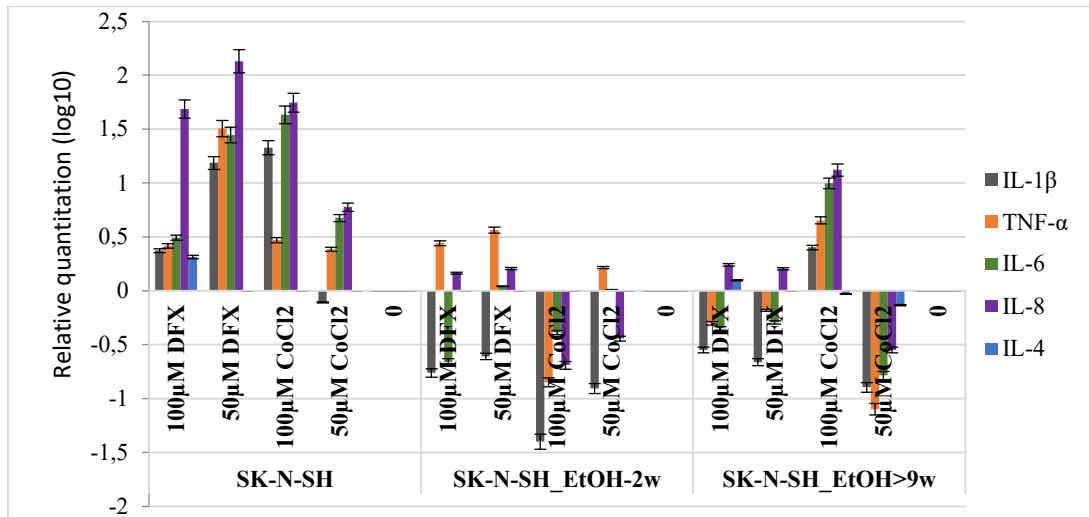


Fig. 7.10. RT PCR Cell stress

The synthesis of cellular stress proteins is shown in Fig. 7.12.

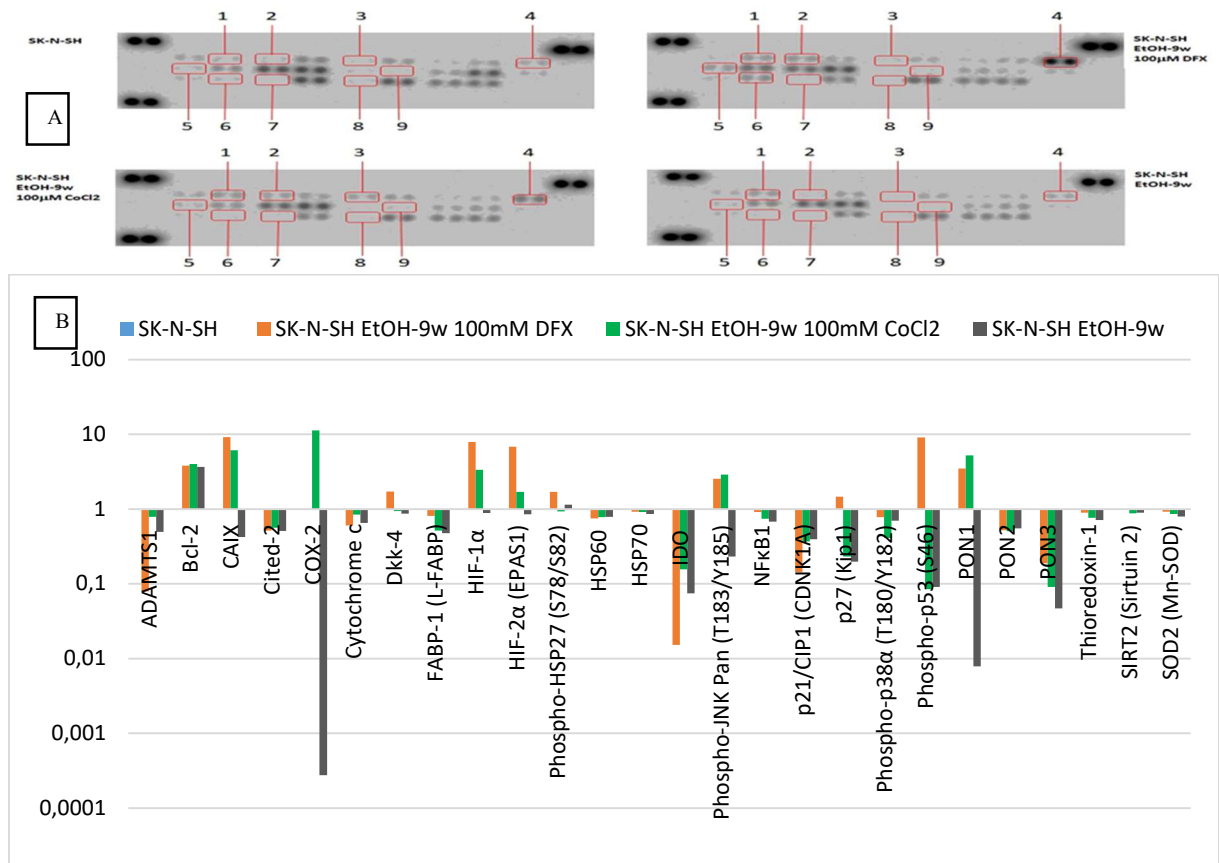


Fig. 7.12. Synthesis of cellular stress proteins under conditions of ethanol exposure and hypoxia (A-view of DotBlot gel and B-graphical representation of logarithmic results)

The gene expressions of the proteins involved in the inflammasome (pyroptosis) are shown in Fig. 7.13.

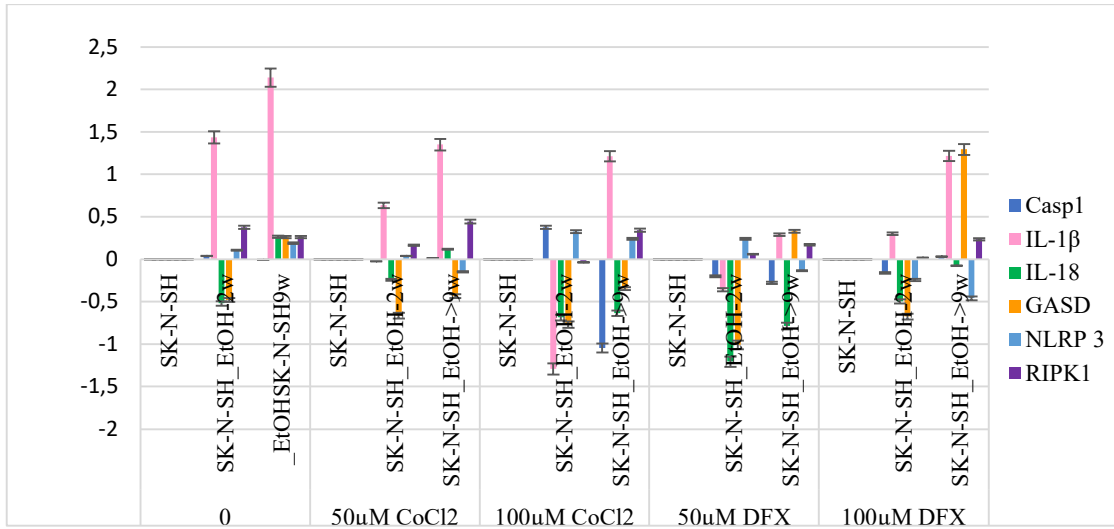


Fig. 7.13. Expression of genes involved in pyroptosis

Gene expressions of carbohydrate metabolism proteins are shown in Fig. 7.16

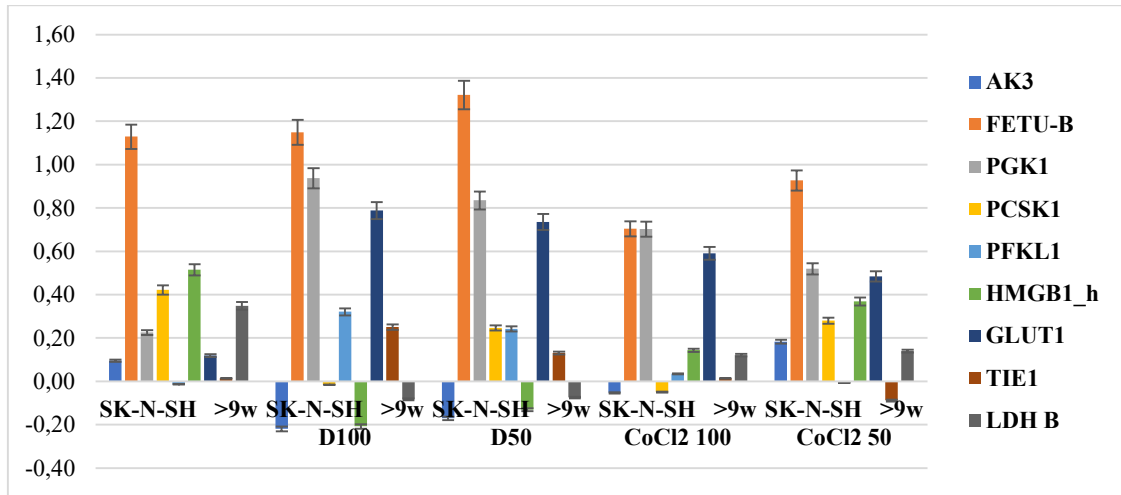
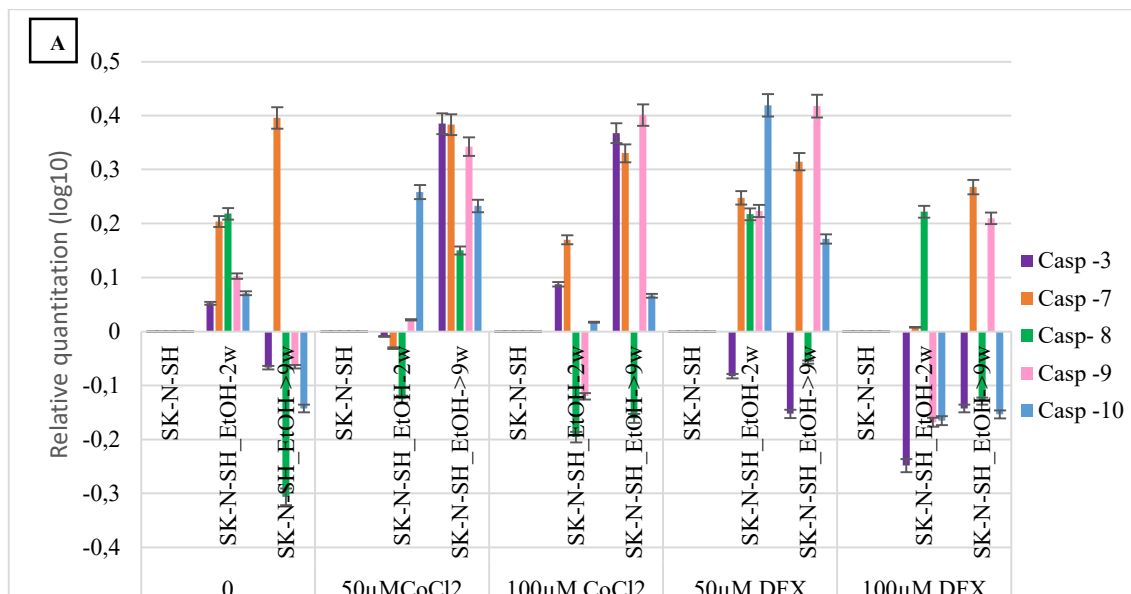


Fig. 7.16. RT PCR Carbohydrate metabolism

Gene expression of apoptotic proteins is shown in Fig. 7.17.



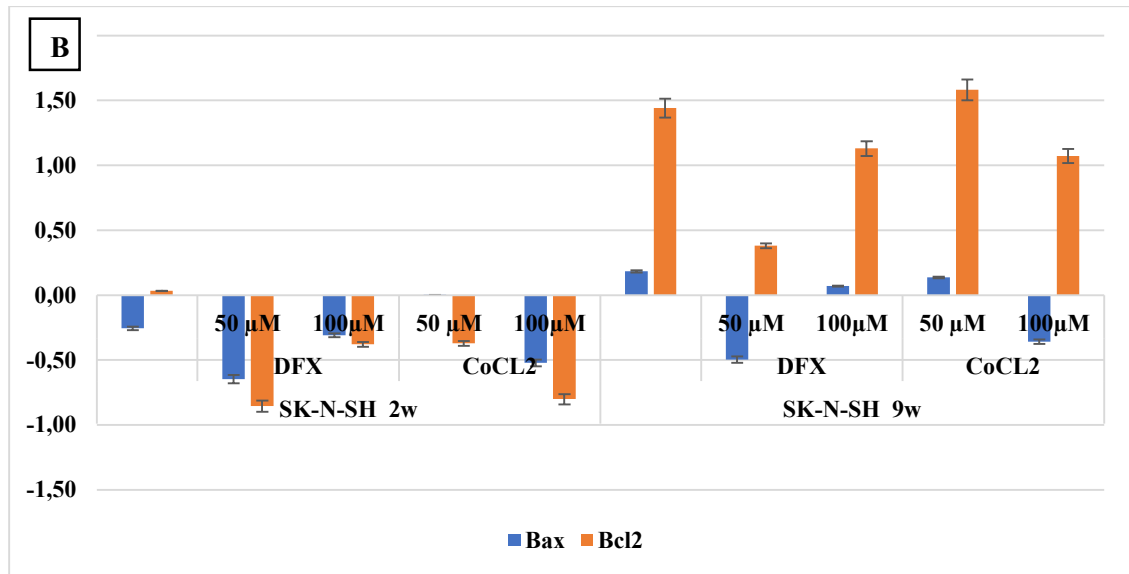


Fig. 7.17. Analysis of gene expression of apoptotic caspases -3, -7, -8, -9, -10 (A) and variation of BAX/Bcl-2 genes (proapoptotic/antiapoptotic) (B)

Apoptotic protein synthesis is depicted in Fig. 7.18.

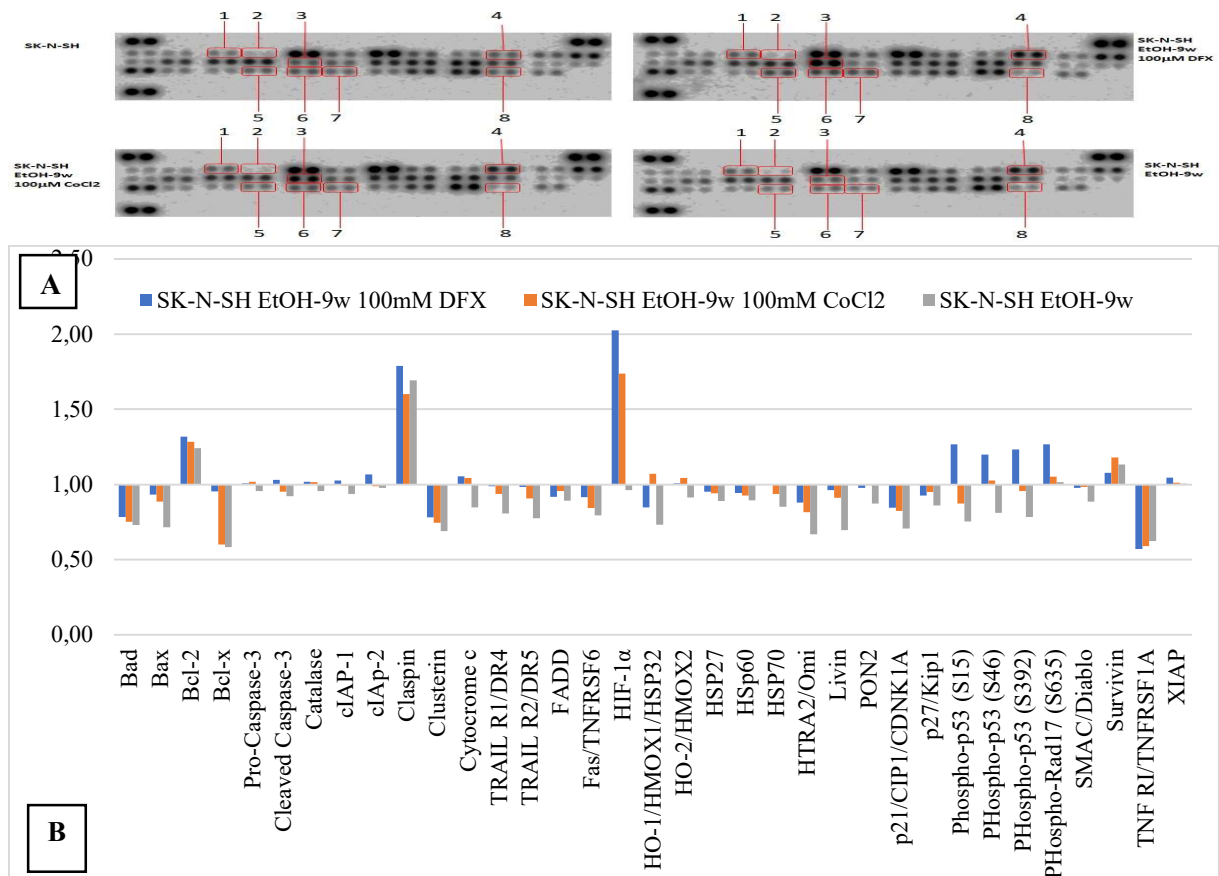


Fig. 7.18. Protein synthesis in apoptosis: A- visualized on experimental DotBlot gel and B- plotted

Gene expression of proteins involved in embryonic signaling pathways is shown in Fig. 7.21.

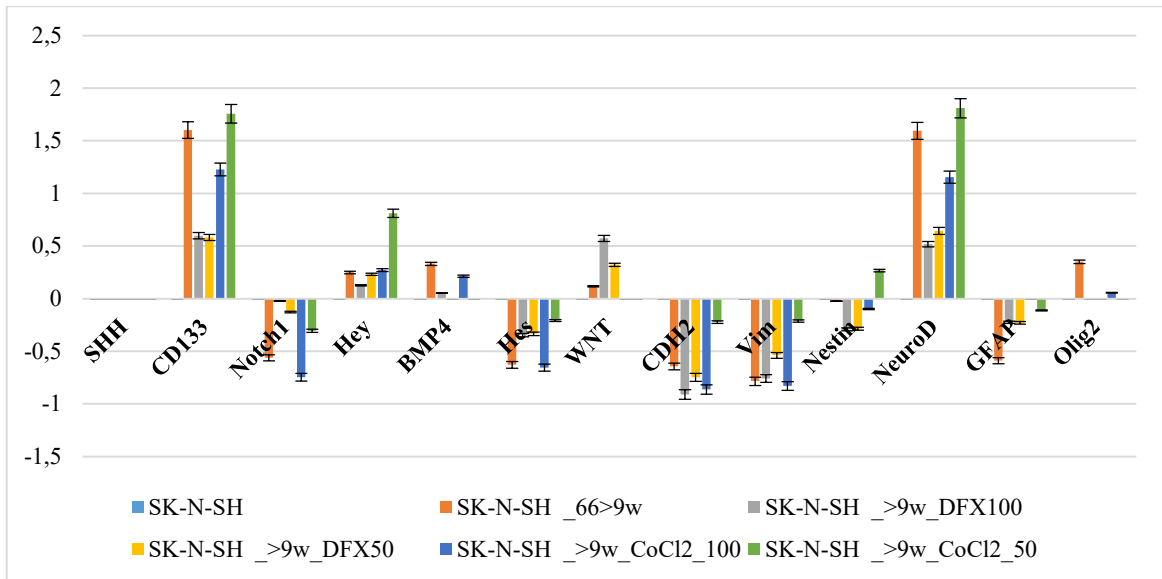


Fig. 7.21. Gene expression analysis of proteins involved in embryonic signaling pathways

Flow cytometry assessment of the cell cycle is shown in Fig. 7.23.

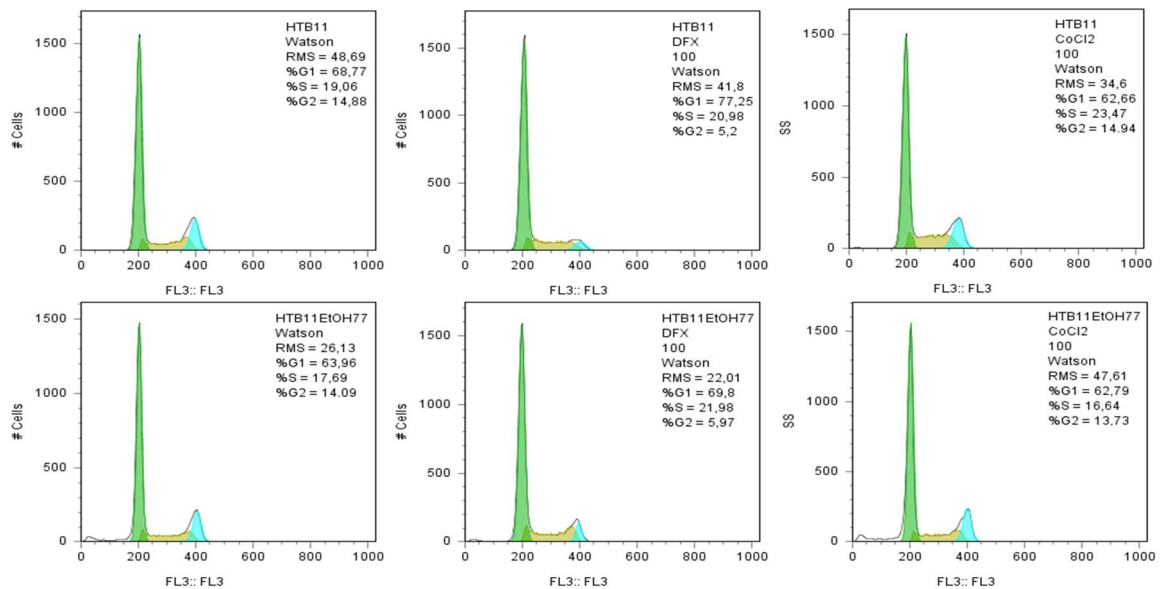


Fig.7.23. Comparative evaluation of the cell cycle

### 8. Study 3: Evaluation of neural cells from primary cell cultures

An important limitation of this PhD is given by the impossibility of carrying out experiments on normal human nerve cells, which is why we made experimental models on human cells from accredited tumor lines. However, out of the desire to follow the cellular behavior under normal conditions, we performed an experiment on neural (glial) cells collected from a newborn rat (of the Wistar species) (following all the recommendations for working on animals) in which we evaluated murine TNF- $\alpha$  and IL-6 protein synthesis [267].

We note the ELISA results of TNF- $\alpha$  and IL-6 protein synthesis in primary cell cultures in Fig. 8.5.

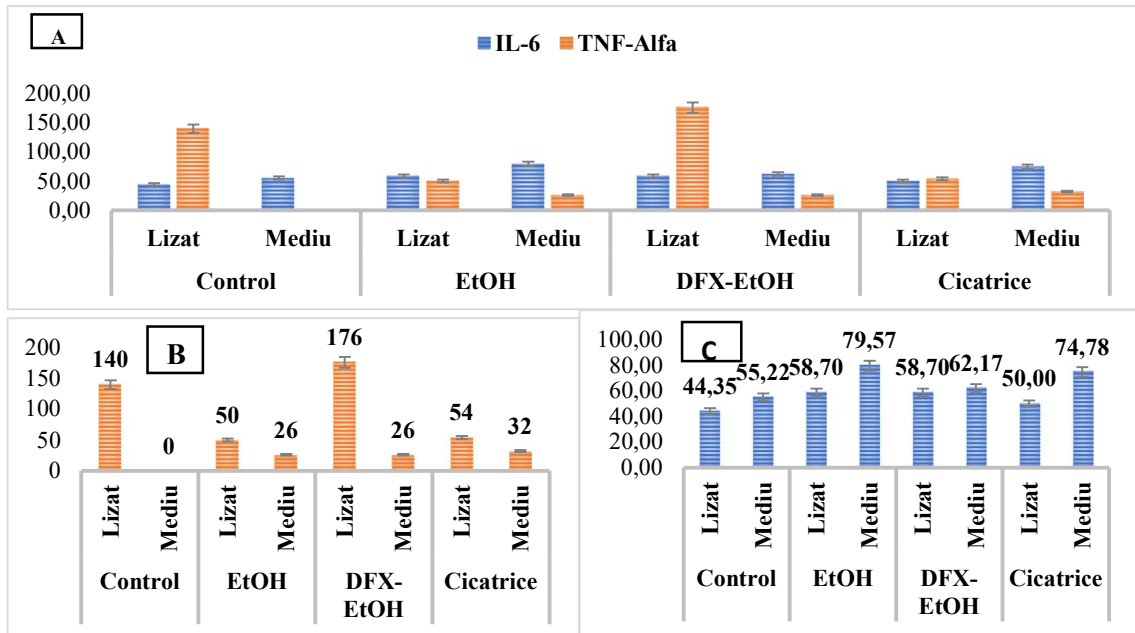


Fig. 8.5. Protein synthesis of TNF- $\alpha$  (B) and IL-6 (C) in gliocytes (in cell lysate and culture medium) (A)

In Fig. 8.6. the values of IL-6 and TNF- $\alpha$  protein synthesis in the studied CSF samples are shown.

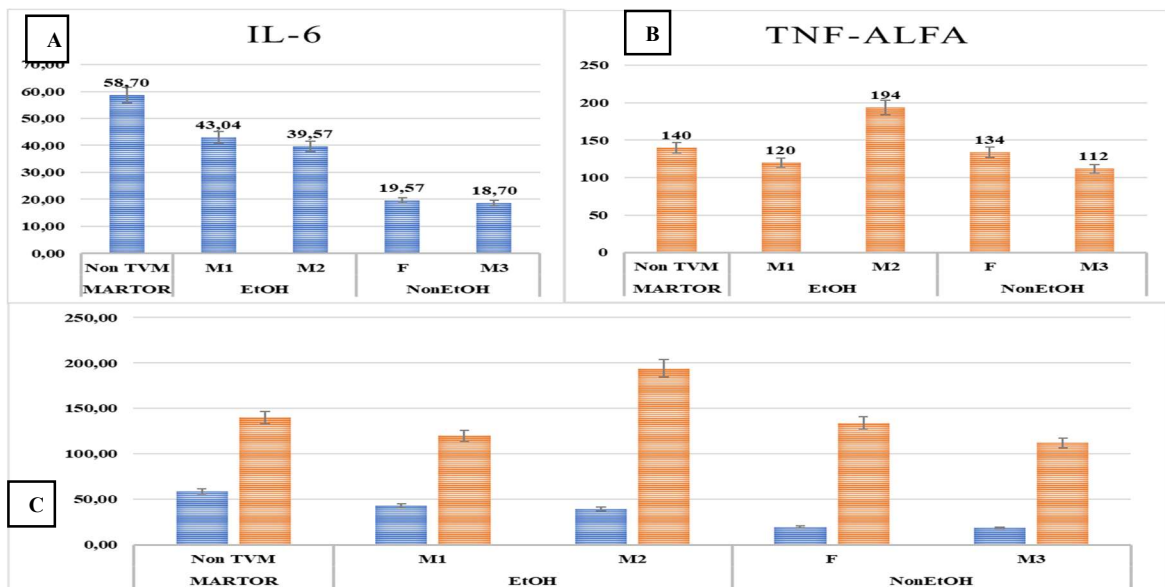


Fig. 8.6. Protein synthesis of TNF- $\alpha$  (B) and IL-6 (A) in CSF (C)

## 9. Conclusions and personal contributions

The complexity of the body's reactions is based on the variety of molecular biology mechanisms and processes that characterize the cells (including) in the spinal cord.

So, the clinical evolution can sometimes be contradictory and surprising in relation to the level of knowledge we have at a given time. Thus, we can reaffirm that ethyl alcohol is a toxic that should not be neglected, because it affects the human body in its entirety and the relationship between ethanol and the nervous system is complex, taking into account the huge neural capacity for adaptation [278], through a plasticity that is far from to be clarified [279] [280]. As we already showed in the introduction: compared to various aggressive elements, at the level of the neurax, complicated and often inter-conditioned elements are formed (which makes it very often extremely difficult, if not impossible, to draw a clear border between the detrimental and the protective) of defense or/ and repairs. Given the still unsatisfactory results regarding the healing possibilities of spinal cord injuries, spinal cord trauma is still a challenge for the medical world, leaving ample room for research, interpretation and progress. Medical recovery is a specialty that intends to reintegrate (familial/social and professional) the patient affected by dysfunctions of various etiologies; and vertebral-medullary trauma occupies a special place in the patient's efforts to return to the previous SCI state.

For all these reasons, we considered useful the research (both from a retrospective statistical point of view, as well as of the intimate mechanisms (genomolecular and cellular) of observational data and objective evaluation quantified, through our clinical experience, which showed (first) a status less severe neuro-dysfunctional and (respectively) a better motor recovery process (in the acute, subacute and chronic stages) in patients with chronic alcoholism compared to those who do not abuse ethanol. Specifically, the retrospective clinical study of 1057 patients with SCI in the acute, subacute stages admitted to the RNM department of SCUBA, over a period of about 15 years, showed (with statistical significance) a better motor status for patients chronic ethanol users, such as and superior performance in consecutive sensory recovery processes.

In our doctoral approach, considering the working hypothesis, we also included prospective fundamental research elements by evaluating the impact of the direct and hypoxic traumatic model on cells exposed to ethanol impregnation and in primary neural tumor and glial cultures. To the extent of technical possibilities and inevitable related limitations,

observational studies were performed on neuroblastoma cell cultures (SK-N-SH) both subjected to chronic ethanol impregnation and without such treatment and (respectively) molecular biology comparisons between reactions of experimental cells (treated or not with ethyl alcohol) to hypoxemic conditions induced by DFX and CoCl<sub>2</sub>.

We also prospectively evaluated (depending on the economic, inherent limitations) some pro-inflammatory and pro-apoptotic molecules in patients with damaged dura mater after SCI, in whom it was thus possible to collect CSF.

We also followed the molecular dynamics following hypoxia and scratch cell injury model. Microscopic analysis of post-traumatic (scratch) lesion status revealed no difference between resilient and (respectively) hypoxic behavior.

The experimental induction of cellular hypoxia as a model of trauma of the studied neurons caused a decrease in HIF-1 $\alpha$  proteolysis, although its gene expression did not increase, followed by trends for: decrease in neurodegeneration and neurotoxicity (by favoring serotonin synthesis produced by inhibition of IDO protein synthesis) , functional improvement of the response/metabolic-functional resilience of neurons (by increasing CA9 and HIF-1 $\beta$ ), neuroprotection (by inhibiting NF-kB gene expression), anti-inflammatory action (by stimulating TGF- $\beta$  gene expression), antiapoptotic (by stimulating the synthesis proteins HMOX1 and HMOX2), vascular regeneration and remodeling (by stimulating VEGF and FLT1 gene expression), synaptic remodeling/neuroplasticity (by stimulating PCDH12 gene expression).

Cellular stress produced by experimental neuronal traumatic models determined the following effects: antioxidant (by stimulating catalase synthesis and by decreasing ROS production, SOD2 and clusterin synthesis), adaptive regulation of energy metabolism (by decreasing HSP60 synthesis), neuroprotective (by stimulating caspase synthesis, HSP27 phosphorylation), anti-inflammatory (by inhibiting IL-1 $\beta$ , TNF- $\alpha$ , IL-6, IL-8 gene expression), antiapoptotic (by inhibiting TNF- $\alpha$  gene expression and decreasing the protein synthesis of HSP70, clusterin, thioredoxin-1), anti-scarring (by inhibiting the expression of the IL-6 gene), inhibitor of nerve regeneration as a defensive response (by inhibiting the expression of the IL-4 gene), restoring nerve endings (by improving the protein synthesis of sirtuin 2), inhibiting of axonal dieback phenomena (by stimulating JNK phosphorylation).

The metabolic changes in experimental neural cells exposed to the hypoxic trauma model determined the following effects: energy depletion (through the inhibition of AK3 gene expression) and lactic acid increase (with a decrease in its dehydrogenation to pyruvic acid), increased insulin proteolytic activity (by stimulating the gene expression of FETU-B), by improving the efficiency of glucose utilization (by stimulating the expression of TIE1), by favoring the intracellular penetration of glucose (by stimulating the expression of the GLUT1 gene) and vascular remodeling (by stimulating the expression of TIE1).

The inflammatory response of experimental neurons exposed to the cell trauma model (by hypoxia) determined the following effects (mediated by the inflammasome): antioxidant (by decreasing the protein synthesis of PON2 and PON3), anti-inflammatory (by inhibiting: caspase-1 gene expression, gene expression NLPR3 after treatment with DFX 50/100 $\mu$ M and 50 $\mu$ M ClCo<sub>2</sub>, of RIPK1 gene expression following treatment with DFX 50/100 $\mu$ M; by decreasing: synthesis of COX2 and NF-kB1, PON2, PON3; of p-38 $\alpha$  phosphorylation), decreasing pyroptosis (by inhibiting gasdermin gene expression), cell survival/biological resistance (by inhibiting gasdermin gene expression).

Regarding programmed cell death by apoptosis, treatment of cell cultures with ethanol inhibits the expression of the proapoptotic gene Bax (toxic mitochondrial) in a manner corresponding to the duration of alcohol exposure, and the inhibition of Bcl-2 expression appears to vary inversely with the duration of ethanol exposure. Analysis of apoptotic protein synthesis shows a correspondence between gene expression and protein synthesis of Bax and Bcl-2, with the inhibition of the apoptotic process augmented by additional hypoxic stress (Bcl-2 synthesis being more increased under hypoxemic conditions). Protein synthesis controlled by the Bax gene is most strongly inhibited in SK-N-SH 9w cultures, while Bcl-2 protein synthesis is stimulated under these conditions; fact that demonstrates an antiapoptotic effect of ethyl alcohol in close relation with the duration of the treatment. Dynamics of Bad protein synthesis show how chronic ethanol treatment inhibits apoptosis more than occurs in the presence of cellular hypoxia. The dynamics of Bcl-x protein synthesis show how apoptotic damage (induced by chronic cellular exposure to ethanol) is inhibited (to a lesser extent) also in the case of induced hypoxic suffering, with a possible more pronounced survival tendency in the situation of long-term alcohol treatment ethyl. The experimental values of procaspase-3 show an antiapoptotic tendency manifested in cells grown in culture media with added ethanol, a tendency that is canceled in hypoxic conditions. The cellular response regarding the synthesis of active caspase-3, cIAP-1 and cIAP-2 shows a decrease

in apoptosis in cells chronically treated with ethanol, an apoptotic decrease that seems to maintain its trend (but to a lesser extent) also in acute hypoxic conditions through treatment with cobalt chloride. Cytochrome C protein synthesis corresponds to the antiapoptotic effect of chronic exposure to ethyl alcohol, a trend that decreases under superadded hypoxic conditions. Synthesis of TRAIL receptors (TNF- $\alpha$  family members): TRAILR1 (DR4) and TRAILR2 (DR5) is inhibited in SK-N-SH 9w cell cultures, a trend that is also maintained under hypoxic conditions. FADD synthesis is inhibited in SK-N-SH 9w cultures, a fact that is maintained (but to a lesser extent) also in the conditions of hypoxemic stress produced by deferoxamine and cobalt chloride. These experimental findings show that chronic exposure to ethanol has antiapoptotic effects, manifested since the initiation of the extrinsic apoptotic pathway. Fas synthesis is inhibited in SK-N-SH cultures grown with chronic exposure to ethanol, a trend that is maintained (but to a lesser extent) also under acute hypoxic conditions produced by cobalt chloride and deferoxamine. OMI/HTRA2 synthesis is inhibited in cell cultures chronically treated with ethanol, a trend that is maintained (but to a lesser extent) when exposed to hypoxic conditions. Livin, a protein belonging to the inhibitor of apoptosis family, is decreased both in SK-N-SH cultures chronically exposed to ethanol and in additional hypoxic conditions produced by treatment with deferoxamine and cobalt chloride, stimulating the apoptotic process. Related to P21 protein synthesis, it was inhibited in case of chronic exposure to ethanol and increased in hypoxic superadded conditions showing that, in hypoxia of cell cultures treated with ethanol, the cell cycle occurs at a slower rate, while the spatial dynamics of the extensions neuronal cell appears to be more active. The literature states that S15 phosphorylation of p53 shows the tendency for neuronal metabolic improvement, with stimulation of neural stem cells, which was also observed following deferoxamine-induced hypoxia. As we have previously shown, data from the literature state that, under stressful conditions, S46 phosphorylation of p53 proves the antiapoptotic tendency present in experimental neuroblastoma cells exposed to hypoxemic treatments. Phosphorylation at serine 392 of p53 has an inhibitory effect on cell development (according to data from the literature referred to previously), and its analysis in cells chronically treated with ethanol revealed better cell development, inclusively compared to acute hypoxic situations. Phosphorylation at serine 635 of Rad17 shows that the stress of prolonged ethanol exposure, augmented by hypoxic conditions slows cell division, due to a tighter control of the cell cycle to prevent genetic errors. Smac/DIABOLO protein synthesis shows a more pronounced antiapoptotic effect in SK-N-SH cultures chronically treated with ethanol, compared to SK-N-SH cultures developed in culture medium without ethanol; an effect that

is also maintained in hypoxic conditions. Protein synthesis of Survivin shows that chronic exposure to ethanol has an antiapoptotic effect on SK-N-SH cultures, an effect that is preserved even in hypoxic conditions. TNFR-1 protein synthesis is inhibited in SK-N-SH cells chronically exposed to ethanol, a trend that is also maintained in hypoxic conditions. XIAP protein synthesis shows an antiapoptotic effect that manifests itself under hypoxic conditions.

Intercellular signaling pathways were investigated at the level of gene expression and protein synthesis for SK-N-SH line neurons treated with ethanol and subjected to the hypoxia cell injury model. From this perspective, neuroprotective trends were observed (by stimulating the synthesis of the Dkk-4 protein, decreasing inflammatory phenomena (by inhibiting the expression of the CHI3L1 and LCN2 genes), reducing neuraxial scars (by inhibiting the expression of the LAMA4, TIMP1, vimentin, nestin, cadherin, Olig2, GAFF, SEMA3, signaling molecules but, at least some of them, also involved in scar development), neurite development and vascular remodeling, neurogenesis (by inhibiting SEMA 3 gene expression and stimulating gene expression BMP4), of synaptogenesis (by decreasing protein synthesis of ADAMTS1), of neuroregeneration (by stimulating Wnt gene expression), of favoring (theoretical) the ability to differentiate into neurons or glial cells in the SK-N-SH line (by stimulating gene expression CD133, NeuroD, FABP-1; by stimulating CD133, NeuroD, FABP-1 gene expression; by decreasing Notch, Hey and Hes gene inhibition). The (theoretical) favorable situation of the ability to differentiate into neurons or glial cells for the SK-N-SH cell line (under the above experimental conditions) was not confirmed by the scratch test (histological finding also valid in connection with the observation of stimulation of experimental neuroregeneration previously described).

Cell cycle analysis did not show significant differences between the cell cultures studied, although P27 protein synthesis shows that prolonged ethanol exposure and added hypoxic conditions cause a favorable response in neuronal cell cycle dynamics. We also reiterate among the conclusions, along with the actual technical and (respectively) related economic limitations of this doctoral work, and the fact that a large part of the steps in the fundamental research were carried out on neuroblastoma lines due to the objective need to be able to work for periods long on immortalized cell cultures; on the other hand, of course, there are also important differences in biological behavior, insculpatory reaction to stress/insults of such tumor cells - and non-neoplastic neurons or gliocytes.

Although ethanol excess negatively influences the nervous system [281] [282] [283] including through a pro-oxidant effect [284][285] the problem of the relationship between ethanol and the human nervous system is very complex. It would seem that the neuraxial suffering from chronic alcoholism causes a decrease in the number of neurons and their biological capacities, with the limitation of hyperexcitability, which can also lead to unsustainable metabolic hyperstimulation with extensive apoptoses - apoptosis like/bad apoptoses, processes followed by the establishment (through severe reduction of neurons at the lesional level and the interruption of the supra/intralesional communication substrate) of the neurofunctional deficit: tetraplegia or paraplegia.

Precisely for this reason, we performed another study on primary glial cell cultures from newborn rats, in which inflammation (IL-6) and apoptosis (TNF- $\alpha$ ) were evaluated after hypoxic traumatization of the respective cells exposed to ethanol excessively, with the following design: control cells not exposed to ethanol and not exposed to hypoxia, cells excessively exposed to ethanol, hypoxically traumatized cells (deferroxamine only, ethanol-impregnated and non-ethanol-impregnated) and thermally traumatized. In summary, the study on primary neuraxial cultures shows (in the case of ethanol exposure) a very efficient inflammatory intercellular communication, with an apoptotic maximum only in the case of combined treatment with ethanol and deferroxamine, but with a low level of apoptotic intercellular communication of cells treated with ethanol (alone or in combination with deferroxamine). In another study (pilot) with a very limited number of participants - for bioethical reasons (the collection was done only in patients with broken dura mater and CSF leakage) the examination revealed: the presence of pro-inflammatory and pro-apoptotic tendencies. We observe how the stress of ethanol treatment, supplemented by hypoxia, accentuates inflammatory intercellular communication, without leading to the highest levels of programmed cell death. The examination of CSF from patients with SCI resumes the discussion about the effects of inflammation and cell death in the initial phases after SCI, when we observe proinflammatory and proapoptotic statuses better represented in patients with chronic alcoholism, possibly also as a mechanism of isolation and protection of the injured medullary area.

The results of our experiments seem surprising, both in the clinical-statistical retrospective dimension, and in those found experimentally propensity. Briefly recapitulating: chronic medullary impregnation with ethanol, we found that it has the main "protective" results after aggressions: a reduction of neuro-excitotoxic phenomena after SCI (including through the

inhibitory action of NMDA receptors [286]), a diminished production of ROS ( including, through a possible mitohormesis effect, stimulating the antioxidant effects within the antioxidant response elements: Antioxidant Response Element - ARE [287,288]) so a lower oxidative stress, a decrease in proapoptotic mechanisms. Of course, these results (which try to explain the geno-molecular and cellular support of the status and evolution, clinically and statistically found to be better in chronic alcoholics) do not exhaust, not far from, the vast yet unexplored field in this extreme complex and complicated field of pathology.

We must not forget how important the dose-effect relationship is, even when it comes to molecules with an antioxidant role (an example of this, not related to our topic, is resveratrol: a supportive molecule for cellular resilience to stress and apoptosis , which can become neurotoxic in quantities that exceed certain limits)[289]. At the same time, the discussion is open on the beneficial effects that can be produced by other molecules included in the composition of alcoholic beverages, as well as the influence of the components of these beverages on other physio-pathological molecular pathways at the level of the cells, including the spinal.

The human nervous system possesses endogenous properties (so far very difficult to capitalize) of post-lesion recovery grouped in the modern concept of endogenous defense activity, neuro-biological function systematized in: neurotrophicity, neuroprotection, neuroplasticity, neuro-(synapto-)genesis [290]. Considering the data from the literature (confirmed also by our current studies) we can say that neuraxial lesions (including SCI) are very complex (with a fragile demarcation between neuroprotective and degenerative ARE processes), without there being (so far) a curative treatment of SCI. In this context, disappointingly decomposed, the chronic ethanol impregnation of the spinal cord tissue deserves to be studied exhaustively in order to try to extract some reactive biological components and pathways from an intimate level that, possibly molecularly separated from the harmful ballast of chronic alcohol ingestion, could find their a potentially valuable therapeutic contribution.

Last but not least, we should reiterate the potential therapeutic effects, in SCI, of molecules structurally similar to ethyl alcohol (for example, polyethylene glycol), which produced in some experimental studies favorable neuroprotective effects [291]: in such situations, research would must persevere to decant the beneficial effects from the toxic ones. The studies in this doctoral thesis have consistent elements of originality, considering that so far

we have not encountered research focused on the (including) favorable effects of chronic ethanol consumption on the recuperative, neurofunctional status and evolution of the neuraxial tissue affected by SCI.

## BIBLIOGRAPHY

- [1] Stoica S-I, Ioana T, Gelu O. Influences and consequences resulting in addictions in general and to chronic alcoholism, especially for patients with spinal cord injury n.d. <https://doi.org/10.12680/balneo.2021.432>.
- [2] Cieza A, Kirchberger I, Biering-Srensen F, Baumberger M, Charlifue S, Post MW, et al. ICF Core Sets for individuals with spinal cord injury in the long-term context. *Spinal Cord* 2010;48:305–12. <https://doi.org/10.1038/sc.2009.183>.
- [3] Barclay L, McDonald R, Lentin P. Social and community participation following spinal cord injury: A critical review. *International Journal of Rehabilitation Research* 2015;38:1–19. <https://doi.org/10.1097/MRR.0000000000000085>.
- [4] Stoica SI, Tănase I, Ciobanu V, Onose G. Initial researches on neuro-functional status and evolution in chronic ethanol consumers with recent traumatic spinal cord injury. *J Med Life* 2019;12. <https://doi.org/10.25122/jml-2019-0026>.
- [5] Kjell J, Olson L. Rat models of spinal cord injury: From pathology to potential therapies. *DMM Disease Models and Mechanisms* 2016;9:1125–37. <https://doi.org/10.1242/dmm.025833>.
- [6] Onose G, Anghelescu A, Muresanu DF, Padure L, Haras MA, Co C, et al. REVIEW A review of published reports on neuroprotection in spinal cord injury. *Spinal Cord* 2009;47:716–26. <https://doi.org/10.1038/sc.2009.52>.
- [7] He X, Li Y, Deng B, Lin A, Zhang G, Ma M, et al. The PI3K/AKT signalling pathway in inflammation, cell death and glial scar formation after traumatic spinal cord injury: Mechanisms and therapeutic opportunities. *Cell Proliferation* 2022:e13275. <https://doi.org/10.1111/CPR.13275>.
- [8] Karsy M, Hawryluk G. Modern Medical Management of Spinal Cord Injury. *Current Neurology and Neuroscience Reports* 2019;19. <https://doi.org/10.1007/s11910-019-0984-1>.

- [9] Kwon BK, Tetzlaff W, Grauer JN, Beiner J, Vaccaro AR. Pathophysiology and pharmacologic treatment of acute spinal cord injury. *Spine Journal* 2004;4:451–64. <https://doi.org/10.1016/j.spinee.2003.07.007>.
- [10] Mei X, Danmeng Z, Wang H, Ru Y, Li F, Wang H, et al. Article 926780 (2022) Mechanism of Ferroptosis and Its Role in. *Frontiers in Neurology | WwwFrontiersinOrg* 2022;1:926780. <https://doi.org/10.3389/fneur.2022.926780>.
- [11] Ruiz IA, Squair JW, Phillips AA, Lukac CD, Huang D, Oxciano P, et al. Incidence and Natural Progression of Neurogenic Shock after Traumatic Spinal Cord Injury. *Journal of Neurotrauma* 2018;35:461–6. <https://doi.org/10.1089/neu.2016.4947>.
- [12] Dietz V, Colombo G. Recovery from spinal cord injury-underlying mechanisms and efficacy of rehabilitation. 2004.
- [13] Onose G. AA, et al. Ghid dediagnostic, tratament și reabilitare însuferințe după traumatisme vertebro-medulare. Bucharest: Ed. Universitara “CarolDavila”; 2011.
- [14] Onose G. AA, et al. „Rehabilitation inconditions following spinal cord injurues”. ”Textbookof Neurosurgery”, vol. 2. Editura Medicala, Bucharest: ISBN 978 - 973-39-0720-6; 2011.
- [15] Maynard FM, Bracken MB, Creasey G, Ditunno JF, Donovan WH, Ducker TB, et al. International Standards for Neurological and Functional Classi@cation of Spinal Cord Injury 1997.
- [16] Khorasanizadeh MH, Yousefifard M, Eskian M, Lu Y, Chalangari M, Harrop JS, et al. Neurological recovery following traumatic spinal cord injury: A systematic review and meta-analysis. *Journal of Neurosurgery: Spine* 2019;30:683–99. <https://doi.org/10.3171/2018.10.SPINE18802>.
- [17] Tse CM, Chisholm AE, Lam T, Eng JJ. A systematic review of the effectiveness of task-specific rehabilitation interventions for improving independent sitting and standing function in spinal cord injury. *Journal of Spinal Cord Medicine* 2018;41:254–66. <https://doi.org/10.1080/10790268.2017.1350340>.
- [18] Ilha J, Meireles A, de Freitas GR, do Espírito Santo CC, Machado-Pereira NAMM, Swarowsky A, et al. Overground gait training promotes functional recovery and cortical

- neuroplasticity in an incomplete spinal cord injury model. *Life Sciences* 2019;232. <https://doi.org/10.1016/j.lfs.2019.116627>.
- [19] Sandrow-Feinberg HR, Houlé JD. Exercise after spinal cord injury as an agent for neuroprotection, regeneration and rehabilitation. *Brain Research* 2015;1619:12–21. <https://doi.org/10.1016/j.brainres.2015.03.052>.
- [20] Maher JL, McMillan DW, Nash MS. Exercise and health-related risks of physical deconditioning after spinal cord injury. *Topics in Spinal Cord Injury Rehabilitation* 2017;23:175–87. <https://doi.org/10.1310/sci2303-175>.
- [21] Bilchak JN, Yeakle K, Caron G, Malloy D, Côté MP. Enhancing KCC2 activity decreases hyperreflexia and spasticity after chronic spinal cord injury. *Experimental Neurology* 2021;338. <https://doi.org/10.1016/j.expneurol.2021.113605>.
- [22] Cowan H, Lakra C, Desai M. Autonomic dysreflexia in spinal cord injury. *BMJ* 2020;371:m3596. <https://doi.org/10.1136/bmj.m3596>.
- [23] Sariyer R, De-Simone FI, Donadoni M, Hoek JB, Chang SL, Sariyer IK. Alcohol-Mediated Missplicing of Mcl-1 Pre-mRNA is Involved in Neurotoxicity. *Alcoholism: Clinical and Experimental Research* 2017;41:1715–24. <https://doi.org/10.1111/acer.13474>.
- [24] Smith SM, Garic A, Flentke GR, Berres ME. Neural Crest Development in Fetal Alcohol Syndrome. *Birth Defects Res C Embryo Today* 2014;102:210. <https://doi.org/10.1002/BDRC.21078>.
- [25] Hauser KF, Khurdayan VK, Goody RJ, Nath A, Saria A, Pauly JR. Selective vulnerability of cerebellar granule neuroblasts and their progeny to drugs with abuse liability. *Cerebellum* 2003;2:184. <https://doi.org/10.1080/14734220310016132>.
- [26] Denny L, Coles S, Blitz R. Fetal Alcohol Syndrome and Fetal Alcohol Spectrum Disorders 2017;96.
- [27] Harper C. The Neuropathology of Alcohol-Related Brain Damage. *Alcohol & Alcoholism* 2009;44:136–40. <https://doi.org/10.1093/alcalc/agn102>.
- [28] Rigler SK. *Alcoholism in the Elderly*. vol. 61. 2000.

- [29] Luo J, Chen G, Wei L, Qian H, Lai X, Wang D, et al. Severe Diffuse Axon Injury in Chronic Alcoholic Rat Medulla Oblongata Following a Concussion Blow n.d. <https://doi.org/10.1093/alcalc/agu009>.
- [30] Harper C, Matsumoto I. Ethanol and brain damage. *Current Opinion in Pharmacology* 2005;5:73–8. <https://doi.org/10.1016/j.coph.2004.06.011>.
- [31] Sarc L, Wraber B, Lipnik-Stangelj M. Ethanol and acetaldehyde disturb TNF-alpha and IL-6 production in cultured astrocytes n.d. <https://doi.org/10.1177/0960327110388533>.
- [32] Gandhirajan A, Roychowdhury S, Kibler C, Bauer SR, Nagy LE, Vachharajani V. Ethanol Exposure Attenuates Immune Response in Sepsis via Sirtuin 2 Expression. *Alcoholism: Clinical and Experimental Research* 2021;45:338–50. <https://doi.org/10.1111/acer.14542>.
- [33] Ann Stephens M, Wand G. Stress and the HPA Axis Role of Glucocorticoids in Alcohol Dependence. n.d.
- [34] Barr T, Helms C, Grant K, Messaoudi I. Opposing effects of alcohol on the immune system. *Progress in Neuro-Psychopharmacology and Biological Psychiatry* 2016;65:242–51. <https://doi.org/10.1016/j.pnpbp.2015.09.001>.
- [35] Blesch A, Lu P, Tsukada S, Alto LT, Roet K, Coppola G, et al. CONDITIONING LESIONS BEFORE OR AFTER SPINAL CORD INJURY RECRUIT BROAD GENETIC MECHANISMS THAT SUSTAIN AXONAL REGENERATION: SUPERIORITY TO CAMP-MEDIATED EFFECTS. *Exp Neurol* 2012;235:162–73. <https://doi.org/10.1016/j.expneurol.2011.12.037>.
- [36] Tabakoff B, Hoffman PL. The neurobiology of alcohol consumption and alcoholism: An integrative history. *Pharmacology Biochemistry and Behavior* 2013;113:20–37. <https://doi.org/10.1016/J.PBB.2013.10.009>.
- [37] Altinoz MA, Elmaci İ. Targeting nitric oxide and NMDA receptor-associated pathways in treatment of high grade glial tumors. Hypotheses for nitro-memantine and nitrones. *Nitric Oxide - Biology and Chemistry* 2018;79:68–83. <https://doi.org/10.1016/j.niox.2017.10.001>.
- [38] Stroud MW, Bombardier CH, Dyer JR, Rimmele CT, Esselman PC. Preinjury alcohol and drug use among persons with spinal cord injury: Implications for rehabilitation. *Journal of Spinal Cord Medicine* 2011;34:461–72. <https://doi.org/10.1179/2045772311Y.0000000033>.

- [39] Bárbara-Bataller E, Méndez-Suárez JL, Alemán-Sánchez C, Sánchez-Enríquez J, Sosa-Henríquez M. Change in the profile of traumatic spinal cord injury over 15 years in Spain. *Scandinavian Journal of Trauma, Resuscitation and Emergency Medicine* 2018;26. <https://doi.org/10.1186/s13049-018-0491-4>.
- [40] <https://pubchem.ncbi.nlm.nih.gov/compound/702> n.d.
- [41] C.Nenitescu. *Chimie organica*. București: Ed Regia Autonoma Monitorul Oficial; 2015.
- [42] <https://pubchem.ncbi.nlm.nih.gov/compound/174#section=Top> n.d.
- [43] Holford NHG. *Clinical Pharmacokinetics of Ethanol*. n.d.
- [44] [https://pubchem.ncbi.nlm.nih.gov/compound/1\\_2-Ethanediol](https://pubchem.ncbi.nlm.nih.gov/compound/1_2-Ethanediol) n.d.
- [45] Vincent Chen H-S, Lipton SA, or Stuart Lipton CA. The chemical biology of clinically tolerated NMDA receptor antagonists. *Journal of Neurochemistry* 2006;97:1611–26. <https://doi.org/10.1111/j.1471-4159.2006.03991.x>.
- [46] *Animal Models and the Cognitive Effects of Ethanol - Animal Models of Cognitive Impairment - NCBI Bookshelf* n.d. <https://www.ncbi.nlm.nih.gov/books/NBK2523/> (accessed July 10, 2022).
- [47] *ALCOHOL DEPENDENCE AND HARMFUL ALCOHOL USE - Alcohol-Use Disorders - NCBI Bookshelf* n.d. <https://www.ncbi.nlm.nih.gov/books/NBK65500/> (accessed July 10, 2022).
- [48] Pelham RW, Nix LC, Chavira RE, Cleveland MV, STETSON P. Clinical trial: single-and multiple-dose pharmacokinetics of polyethylene glycol (PEG-3350) in healthy young and elderly subjects n.d. <https://doi.org/10.1111/j.1365-2036.2008.03727.x>.
- [49] Hagen EM. Acute complications of spinal cord injuries. *World Journal of Orthopedics* 2015;6. <https://doi.org/10.5312/wjo.v6.i1.17>.
- [50] Yanagisawa S, Katoh H, Imai T, Nomura S, Watanabe M. The relationship between inflammasomes and the endoplasmic reticulum stress response in the injured spinal cord. *Neuroscience Letters* 2019;705:54–9. <https://doi.org/10.1016/J.NEULET.2019.04.033>.
- [51] Rowland JW, Hawryluk GWJ, Kwon B, Fehlings MG. Current status of acute spinal cord injury pathophysiology and emerging therapies: Promise on the horizon. *Neurosurgical Focus* 2008;25. <https://doi.org/10.3171/FOC.2008.25.11.E2>.

- [52] Bruce Alberts AJJLDMMRKPW. *Molecular Biology of the Cell*. Sixth. New York : 2014.
- [53] Hayashi M, Ueyama T, Nemoto K, Tamaki T, Senba E. Sequential mRNA expression for immediate early genes, cytokines, and neurotrophins in spinal cord injury. *Journal of Neurotrauma* 2000;17. <https://doi.org/10.1089/neu.2000.17.203>.
- [54] Quan HH, Kang KS, Sohn YK, Li M. Tempol reduces injury area in rat model of spinal cord contusion injury through suppression of iNOS and COX-2 expression. *Neurol Sci* 2013;34:1621–8. <https://doi.org/10.1007/S10072-013-1295-Y>.
- [55] López-Vales R, García-Alías G, Guzmán-Lenis MS, Forés J, Casas C, Navarro X, et al. Effects of COX-2 and iNOS Inhibitors Alone or in Combination With Olfactory Ensheathing Cell Grafts After Spinal Cord Injury. vol. 31. n.d.
- [56] Alizadeh A, Dyck SM, Karimi-Abdolrezaee S. Traumatic Spinal Cord Injury: An Overview of Pathophysiology, Models and Acute Injury Mechanisms. *Frontiers in Neurology* 2019;10:282. <https://doi.org/10.3389/FNEUR.2019.00282>.
- [57] Hall ED, Springer JE. *Neuroprotection and Acute Spinal Cord Injury: A Reappraisal* n.d.
- [58] Hernandez-Gerez E, Fleming IN, Parson SH. A role for spinal cord hypoxia in neurodegeneration n.d. <https://doi.org/10.1038/s41419-019-2104-1>.
- [59] Wu D, Yotnda P. Induction and Testing of Hypoxia in Cell Culture. *Journal of Visualized Experiments : JoVE* 2011:2899. <https://doi.org/10.3791/2899>.
- [60] Maxwell PH, Wlesener MS, Chang GW, Clifford SC, Vaux EC, Cockman ME, et al. The tumour suppressor protein VHL targets hypoxia-inducible factors for oxygen-dependent proteolysis. *Nature* 1999;399:271–5. <https://doi.org/10.1038/20459>.
- [61] Takacova M, Barathova M, Zatovicova M, Golias T, Kajanova I, Jelenska L, et al. Carbonic Anhydrase IX—Mouse versus Human. *International Journal of Molecular Sciences* 2020;21. <https://doi.org/10.3390/IJMS21010246>.
- [62] Maurer MH, Tripps WKC, Feldmann RE, Kuschinsky W. Expression of vascular endothelial growth factor and its receptors in rat neural stem cells. *Neuroscience Letters* 2003;344:165–8. [https://doi.org/10.1016/S0304-3940\(03\)00407-5](https://doi.org/10.1016/S0304-3940(03)00407-5).
- [63] Ratcliffe PJ. HIF-1 and HIF-2: working alone or together in hypoxia? *The Journal of Clinical Investigation* 2007;117:862–5. <https://doi.org/10.1172/JCI31750>.

- [64] Peyronnard JM, Messier JP, Charron L, Lavoie J, Bergouignan FX, Dubreuil M. Carbonic anhydrase activity in the normal and injured peripheral nervous system of the rat. *Exp Neurol* 1986;93:481–99. [https://doi.org/10.1016/0014-4886\(86\)90169-X](https://doi.org/10.1016/0014-4886(86)90169-X).
- [65] Riley DA, Ellis S, Bain JLW. Ultrastructural cytochemical localization of carbonic anhydrase activity in rat peripheral sensory and motor nerves, dorsal root ganglia and dorsal column nuclei. *Neuroscience* 1984;13:189–206. [https://doi.org/10.1016/0306-4522\(84\)90269-0](https://doi.org/10.1016/0306-4522(84)90269-0).
- [66] Wang B, Samanani N, Roth SH, Archer DP. Spinal Carbonic Anhydrase Contributes to Nociceptive Reflex Enhancement by Midazolam, Pentobarbital, and Propofol. *Anesthesiology* 2003;98:921–7. <https://doi.org/10.1097/00000542-200304000-00019>.
- [67] Parfenova H, Leffler C. Cerebroprotective Functions of HO-2. *Current Pharmaceutical Design* 2008;14. <https://doi.org/10.2174/138161208783597380>.
- [68] Hanna DA, Moore CM, Liu L, Yuan X, Dominic IM, Fleischhacker AS, et al. Heme oxygenase-2 (HO-2) binds and buffers labile ferric heme in human embryonic kidney cells. *Journal of Biological Chemistry* 2022;298:101549. <https://doi.org/10.1016/j.jbc.2021.101549>.
- [69] Dunn LL, Kong SMY, Tumanov S, Chen W, Cantley J, Ayer A, et al. Hmox1 (Heme Oxygenase-1) Protects against Ischemia-Mediated Injury via Stabilization of HIF-1 $\alpha$  (Hypoxia-Inducible Factor-1 $\alpha$ ). *Arteriosclerosis, Thrombosis, and Vascular Biology* 2021;41. <https://doi.org/10.1161/ATVBAHA.120.315393>.
- [70] Jazwa A, Stoszko M, Tomczyk M, Bukowska-Strakova K, Pichon C, Jozkowicz A, et al. HIF-regulated HO-1 gene transfer improves the post-ischemic limb recovery and diminishes TLR-triggered immune responses - Effects modified by concomitant VEGF overexpression. *Vascular Pharmacology* 2015;71. <https://doi.org/10.1016/j.vph.2015.02.011>.
- [71] Salminen LE, Schofield PR, Pierce KD, Bruce SE, Griffin MG, Tate DF, et al. Vulnerability of white matter tracts and cognition to the SOD2 polymorphism: A preliminary study of antioxidant defense genes in brain aging. *Behavioural Brain Research* 2017;329:111. <https://doi.org/10.1016/J.BBR.2017.04.041>.
- [72] Kase BA, Northrup H, Morrison AC, Davidson CM, Goiffon AM, Fletcher JM, et al. Association of Copper-Zinc Superoxide Dismutase (SOD1) and Manganese Superoxide

Dismutase (SOD2) Genes with Non-syndromic Myelomeningocele. *Birth Defects Res A Clin Mol Teratol* 2012;94:762. <https://doi.org/10.1002/BDRA.23065>.

- [73] Barr RK, Bogoyevitch MA. The c-Jun N-terminal protein kinase family of mitogen-activated protein kinases (JNK MAPKs). vol. 33. 2001.
- [74] Yoshimura K, Ueno M, Lee S, Nakamura Y, Sato A, Yoshimura K, et al. C-Jun N-terminal kinase induces axonal degeneration and limits motor recovery after spinal cord injury in mice. *Neuroscience Research* 2011;71:266–77. <https://doi.org/10.1016/J.NEURES.2011.07.1830>.
- [75] Kasuya Y, Umezawa H, Hatano M. Stress-Activated Protein Kinases in Spinal Cord Injury: Focus on Roles of p38. *International Journal of Molecular Sciences* 2018;19. <https://doi.org/10.3390/IJMS19030867>.
- [76] Navarro-zaragoza J, Cuenca-bermejo L, Almela P, Laorden ML, Herrero MT. Could small heat shock protein hsp27 be a first-line target for preventing protein aggregation in parkinson's disease? *International Journal of Molecular Sciences* 2021;22. <https://doi.org/10.3390/ijms22063038>.
- [77] Hecker JG, McGarvey M. Heat shock proteins as biomarkers for the rapid detection of brain and spinal cord ischemia: A review and comparison to other methods of detection in thoracic aneurysm repair. *Cell Stress and Chaperones* 2011;16. <https://doi.org/10.1007/s12192-010-0224-8>.
- [78] Rodriguez A, von Salzen D, Holguin BA, Bernal RA. Complex Destabilization in the Mitochondrial Chaperonin Hsp60 Leads to Disease. *Frontiers in Molecular Biosciences* 2020;7. <https://doi.org/10.3389/fmolb.2020.00159>.
- [79] Xie C, Shen X, Xu X, Liu H, Li F, Lu S, et al. Astrocytic YAP Promotes the Formation of Glia Scars and Neural Regeneration after Spinal Cord Injury. *The Journal of Neuroscience* 2020;40:2644. <https://doi.org/10.1523/JNEUROSCI.2229-19.2020>.
- [80] Noma T. Dynamics of nucleotide metabolism as a supporter of life phenomena. *Journal of Medical Investigation* 2005;52:127–36. <https://doi.org/10.2152/JMI.52.127>.
- [81] Tamai M, Kawano T, Saito R, Sakurai K, Saito Y, Yamada H, et al. Phosphoglycerate kinase deficiency due to a novel mutation (c. 1180A>G) manifesting as chronic hemolytic anemia

in a Japanese boy. *International Journal of Hematology* 2014;393–7. <https://doi.org/10.1007/S12185-014-1615-X>.

- [82] Wilson RB, Solass W, Archid R, Weinreich FJ, Königsrainer A, Reymond MA. Resistance to anoikis in transcoelomic shedding: the role of glycolytic enzymes. *Pleura Peritoneum* 2019;4. <https://doi.org/10.1515/PP-2019-0003>.
- [83] Gao L, Wang C, Qin B, Li T, Xu W, Lenahan C, et al. 6-phosphofructo-2-kinase/fructose-2,6-bisphosphatase Suppresses Neuronal Apoptosis by Increasing Glycolysis and “cyclin-dependent kinase 1-Mediated Phosphorylation of p27 After Traumatic Spinal Cord Injury in Rats. *Cell Transplantation* 2020;29. <https://doi.org/10.1177/0963689720950226>.
- [84] Zheng M, Streck FD, Sc~tt FEM, Seidah ~ N G, Pintar JE. The Developmental Expression in Rat of Proteases Furin, PC1 , PC2, and Carboxypeptidase E: Implications for Early Maturation of Proteolytic Processing Capacity. *The Journal of Neuroscience* 1994;14:4858–73.
- [85] Whetstone WD, Hsu JYC, Eisenberg M, Werb Z, Noble-Haeusslein LJ. Blood-Spinal Cord Barrier After Spinal Cord Injury: Relation to Revascularization and Wound Healing. *J Neurosci Res* 2003;74:227. <https://doi.org/10.1002/JNR.10759>.
- [86] Auvin S. Abnormal white matter: Expanding the GLUT1-D phenotype. *European Journal of Paediatric Neurology* 2018;22:345. <https://doi.org/10.1016/J.EJPN.2018.03.007>.
- [87] Duka T, Anderson SM, Collins Z, Raghanti MA, Ely JJ, Hof PR, et al. SYNAPTOSOMAL LACTATE DEHYDROGENASE ISOENZYME COMPOSITION IS SHIFTED TOWARD AEROBIC FORMS IN PRIMATE BRAIN EVOLUTION. *Brain Behav Evol* 2014;83:216. <https://doi.org/10.1159/000358581>.
- [88] Lu X, Al-Aref R, Zhao D, Shen J, Yan Y, Gao Y. Astrocyte-conditioned medium attenuates glutamate-induced apoptotic cell death in primary cultured spinal cord neurons of rats. *Neurological Research* 2015;37:803–8. <https://doi.org/10.1179/1743132815Y.0000000059>.
- [89] Cañete-Soler R, Reddy KS, Tolan DR, Zhai J. Aldolases A and C Are Ribonucleolytic Components of a Neuronal Complex That Regulates the Stability of the Light-Neurofilament mRNA. *The Journal of Neuroscience* 2005;25:4353. <https://doi.org/10.1523/JNEUROSCI.0885-05.2005>.

- [90] Blackmore MG, Moore DL, Smith RP, Goldberg JL, Bixby JL, Lemmon VP. High Content Screening of Cortical Neurons Identifies Novel Regulators of Axon Growth. *Mol Cell Neurosci* 2010;44:43. <https://doi.org/10.1016/J.MCN.2010.02.002>.
- [91] Ulbrich L, Cozzolino M, Marini ES, Amori I, de Jaco A, Carri MT, et al. Cystatin B and SOD1: Protein-protein interaction and possible relation to neurodegeneration. *Cellular and Molecular Neurobiology* 2014;34:205–13. <https://doi.org/10.1007/s10571-013-0004-y>.
- [92] Daura E, Tegelberg S, Yoshihara M, Jackson C, Simonetti F, Aksentjeff K, et al. Cystatin B-deficiency triggers ectopic histone H3 tail cleavage during neurogenesis. *Neurobiology of Disease* 2021;156. <https://doi.org/10.1016/J.NBD.2021.105418>.
- [93] Okuneva O, Li Z, Körber I, Tegelberg S, Joensuu T, Tian L, et al. Brain inflammation is accompanied by peripheral inflammation in *Cstb*<sup>-/-</sup> mice, a model for progressive myoclonus epilepsy. *Journal of Neuroinflammation* 2016;13. <https://doi.org/10.1186/S12974-016-0764-7>.
- [94] Xue S, Han H, Rui S, Yang M, Huang Y, Zhan B, et al. Serum Fetuin-B Levels Are Elevated in Women with Metabolic Syndrome and Associated with Increased Oxidative Stress 2021. <https://doi.org/10.1155/2021/6657658>.
- [95] Karmilin K, Schmitz C, Kuske M, Körschgen H, Olf M, Meyer K, et al. Mammalian plasma fetuin-B is a selective inhibitor of ovastacin and meprin metalloproteinases. *Scientific Reports* 2019;9. <https://doi.org/10.1038/S41598-018-37024-5>.
- [96] Peter A, Kovarova M, Staiger H, Machann J, Schick F, Königsrainer A, et al. The hepatokines fetuin-A and fetuin-B are upregulated in the state of hepatic steatosis and may differently impact on glucose homeostasis in humans. *American Journal of Physiology - Endocrinology and Metabolism* 2018;314:E266–73. <https://doi.org/10.1152/AJPENDO.00262.2017>.
- [97] Kuniyama T, Sasaki S, Shiiya N, Ishikura H, Kawarada Y, Matsukawa A, et al. Lazaroid reduces production of IL-8 and IL-1 receptor antagonist in ischemic spinal cord injury. *The Annals of Thoracic Surgery* 2000;69:792–8. [https://doi.org/10.1016/S0003-4975\(99\)01413-7](https://doi.org/10.1016/S0003-4975(99)01413-7).
- [98] Dalkilic T, Fallah N, Noonan VK, Salimi Elizei S, Dong K, Belanger L, et al. Predicting Injury Severity and Neurological Recovery after Acute Cervical Spinal Cord Injury: A

Comparison of Cerebrospinal Fluid and Magnetic Resonance Imaging Biomarkers. *Journal of Neurotrauma* 2018;35:435–45. <https://doi.org/10.1089/neu.2017.5357>.

- [99] Gordon R, Albornoz EA, Christie DC, Langley MR, Kumar V, Mantovani S, et al. Inflammasome inhibition prevents  $\alpha$ -synuclein pathology and dopaminergic neurodegeneration in mice. *Science Translational Medicine* 2018;10. <https://doi.org/10.1126/scitranslmed.aah4066>.
- [100] Mortezaee K, Khanlarkhani N, Beyer C, Zendedel A. Inflammasome: Its role in traumatic brain and spinal cord injury. *Journal of Cellular Physiology* 2018;233. <https://doi.org/10.1002/jcp.26287>.
- [101] de Vasconcelos NM, Lamkanfi M. Recent insights on inflammasomes, gasdermin pores, and pyroptosis. *Cold Spring Harbor Perspectives in Biology* 2020;12. <https://doi.org/10.1101/cshperspect.a036392>.
- [102] Voet S, Srinivasan S, Lamkanfi M, Loo G. Inflammasomes in neuroinflammatory and neurodegenerative diseases. *EMBO Molecular Medicine* 2019;11. <https://doi.org/10.15252/emmm.201810248>.
- [103] Swanson K v., Deng M, Ting JPY. The NLRP3 inflammasome: molecular activation and regulation to therapeutics. *Nature Reviews Immunology* 2019;19. <https://doi.org/10.1038/s41577-019-0165-0>.
- [104] Lammert CR, Frost EL, Bellinger CE, Bolte AC, Mckee CA, Hurt ME, et al. AIM2 inflammasome surveillance of DNA damage shapes neurodevelopment HHS Public Access n.d. <https://doi.org/10.1038/s41586-020-2174-3>.
- [105] Man SM, Karki R, Kanneganti TD. Molecular mechanisms and functions of pyroptosis, inflammatory caspases and inflammasomes in infectious diseases. *Immunol Rev* 2017;277:61. <https://doi.org/10.1111/IMR.12534>.
- [106] Kelley N, Jeltema D, Duan Y, He Y. The NLRP3 inflammasome: An overview of mechanisms of activation and regulation. *International Journal of Molecular Sciences* 2019;20. <https://doi.org/10.3390/ijms20133328>.
- [107] Prendergast GC, Malachowski WJ, Mondal A, Scherle P, Muller AJ. Indoleamine 2,3-Dioxygenase and Its Therapeutic Inhibition in Cancer. *Int Rev Cell Mol Biol* 2018;336:175. <https://doi.org/10.1016/BS.IRCMB.2017.07.004>.

- [108] Barreto FS, Filho AJMC, de Araújo MCCR, de Moraes MO, de Moraes MEA, Maes M, et al. Tryptophan catabolites along the indoleamine 2,3-dioxygenase pathway as a biological link between depression and cancer. *Behavioural Pharmacology* 2018;29:165–80. <https://doi.org/10.1097/FBP.0000000000000384>.
- [109] Sakurai K, Zou J-P, Tschetter JR, Ward JM, Shearer GM. Effect of indoleamine 2,3-dioxygenase on induction of experimental autoimmune encephalomyelitis. n.d.
- [110] Huang YS, Ogbechi J, Clanchy FI, Williams RO, Stone TW. IDO and Kynurenine Metabolites in Peripheral and CNS Disorders. *Frontiers in Immunology* 2020;11:388. <https://doi.org/10.3389/FIMMU.2020.00388>.
- [111] Godin-Ethier J, Hanafi LA, Duvignaud JB, Leclerc D, Lapointe R. IDO expression by human B lymphocytes in response to T lymphocyte stimuli and TLR engagement is biologically inactive. *Molecular Immunology* 2011;49:253–9. <https://doi.org/10.1016/J.MOLIMM.2011.08.017>.
- [112] Lam CS, Li JJ, Tipoe GL, Youdim MBH, Fung ML. Monoamine oxidase A upregulated by chronic intermittent hypoxia activates indoleamine 2,3-dioxygenase and neurodegeneration. *PLoS ONE* 2017;12. <https://doi.org/10.1371/journal.pone.0177940>.
- [113] Costa LG, de Laat R, Dao K, Pellacani C, Cole TB, Furlong CE. Paraoxonase-2 (PON2) in brain and its potential role in neuroprotection. *NeuroToxicology* 2014;43. <https://doi.org/10.1016/j.neuro.2013.08.011>.
- [114] Manco G, Porzio E, Carusone TM. Human paraoxonase-2 (Pon2): Protein functions and modulation. *Antioxidants* 2021;10. <https://doi.org/10.3390/antiox10020256>.
- [115] Fiorentino L, Stehlik C, Oliveira V, Ariza ME, Godzik A, Reed JC. A novel PAAD-containing protein that modulates NF- $\kappa$ B induction by cytokines tumor necrosis factor- $\alpha$  and interleukin-1 $\beta$ . *Journal of Biological Chemistry* 2002;277:35333–40. <https://doi.org/10.1074/jbc.M200446200>.
- [116] Smits VAJ, Cabrera E, Freire R, Gillespie DA. Claspin - checkpoint adaptor and DNA replication factor. *FEBS J* 2019;286:441–55. <https://doi.org/10.1111/FEBS.14594>.
- [117] Zinkie S, Gentil BJ, Minotti S, Durham HD. Expression of the protein chaperone, clusterin, in spinal cord cells constitutively and following cellular stress, and upregulation by treatment

with Hsp90 inhibitor. *Cell Stress and Chaperones* 2013;18. <https://doi.org/10.1007/s12192-013-0427-x>.

- [118] Wright MC, Mi R, Connor E, Reed N, Vyas A, Alspalter M, et al. Novel roles for osteopontin and clusterin in peripheral motor and sensory axon regeneration. *Journal of Neuroscience* 2014;34. <https://doi.org/10.1523/JNEUROSCI.3822-13.2014>.
- [119] Shafie INF, McLaughlin M, Burchmore R, Lim MAA, Montague P, Johnston PEJ, et al. The chaperone protein clusterin may serve as a cerebrospinal fluid biomarker for chronic spinal cord disorders in the dog. *Cell Stress and Chaperones* 2014;19. <https://doi.org/10.1007/s12192-013-0457-4>.
- [120] Fang P, Pan HC, Lin SL, Zhang WQ, Rauvala H, Schachner M, et al. HMGB1 contributes to regeneration after spinal cord injury in adult zebrafish. *Mol Neurobiol* 2014;49:472–83. <https://doi.org/10.1007/S12035-013-8533-4>.
- [121] Galluzzi L, Vitale I. Molecular mechanisms of cell death: recommendations of the Nomenclature Committee on Cell Death 2018. *Cell Death & Differentiation* 2018;25:486–541. <https://doi.org/10.1038/s41418-017-0012-4>.
- [122] Khoury MK, Gupta K, Franco SR, Liu B. Necroptosis in the Pathophysiology of Disease. *American Journal of Pathology* 2020;190. <https://doi.org/10.1016/j.ajpath.2019.10.012>.
- [123] Fricker M, Tolkovsky AM, Borutaite V, Coleman M, Brown GC. Neuronal cell death. *Physiological Reviews* 2018;98. <https://doi.org/10.1152/physrev.00011.2017>.
- [124] Frank D, Vince JE. Pyroptosis versus necroptosis: similarities, differences, and crosstalk. *Cell Death and Differentiation* 2019;26. <https://doi.org/10.1038/s41418-018-0212-6>.
- [125] Long JS, Ryan KM. New frontiers in promoting tumour cell death: Targeting apoptosis, necroptosis and autophagy. *Oncogene* 2012;31. <https://doi.org/10.1038/onc.2012.7>.
- [126] Pistritto G, Trisciuglio D, Ceci C, Alessia Garufi, D’Orazi G. Apoptosis as anticancer mechanism: Function and dysfunction of its modulators and targeted therapeutic strategies. *Aging* 2016;8. <https://doi.org/10.18632/aging.100934>.
- [127] Xu X, Lai Y, Hua ZC. Apoptosis and apoptotic body: Disease message and therapeutic target potentials. *Bioscience Reports* 2019;39. <https://doi.org/10.1042/BSR20180992>.

- [128] Paul A, Krelin Y, Arif T, Jeger R, Shoshan-Barmatz V. A New Role for the Mitochondrial Pro-apoptotic Protein SMAC/Diablo in Phospholipid Synthesis Associated with Tumorigenesis. *Molecular Therapy* 2018;26. <https://doi.org/10.1016/j.ymthe.2017.12.020>.
- [129] Fung S, Knoefel WT, Krieg A. Clinicopathological and prognostic significance of inhibitor of apoptosis protein (IAP) family members in lung cancer: A meta-analysis. *Cancers (Basel)* 2021;13. <https://doi.org/10.3390/cancers13164098>.
- [130] Wheatley SP, Altieri DC. Survivin at a glance. *Journal of Cell Science* 2019;132. <https://doi.org/10.1242/jcs.223826>.
- [131] Altieri B, Sbiera S, Casa S della, Weigand I, Wild V, Steinhauer S, et al. Livin/BIRC7 expression as malignancy marker in adrenocortical tumors. *Oncotarget* 2017;8:9323. <https://doi.org/10.18632/ONCOTARGET.14067>.
- [132] Gonzalez YR, Zhang Y, Behzadpoor D, Cregan S, Bamforth S, Slack RS, et al. CITED2 Signals through Peroxisome Proliferator-Activated Receptor- $\gamma$  to Regulate Death of Cortical Neurons after DNA Damage. *The Journal of Neuroscience* 2008;28:5559. <https://doi.org/10.1523/JNEUROSCI.1014-08.2008>.
- [133] Oda K, Arakawa H, Tanaka T, Matsuda K, Tanikawa C, Mori T, et al. p53AIP1, a potential mediator of p53-dependent apoptosis, and its regulation by ser-46-phosphorylated p53. *Cell* 2000;102:849–62. [https://doi.org/10.1016/S0092-8674\(00\)00073-8](https://doi.org/10.1016/S0092-8674(00)00073-8).
- [134] Domínguez-Bautista JA, Acevo-Rodríguez PS, Castro-Obregón S. Programmed Cell Senescence in the Mouse Developing Spinal Cord and Notochord. *Frontiers in Cell and Developmental Biology* 2021;9. <https://doi.org/10.3389/fcell.2021.587096>.
- [135] Imamura T, Uesaka M, Nakashima K. Epigenetic setting and reprogramming for neural cell fate determination and differentiation. *Philosophical Transactions of the Royal Society B: Biological Sciences* 2014;369. <https://doi.org/10.1098/rstb.2013.0511>.
- [136] Li S, Leshchyn'ska I, Chernyshova Y, Schachner M, Sytnyk V. The Neural Cell Adhesion Molecule (NCAM) Associates with and Signals through p21-Activated Kinase 1 (Pak1). *The Journal of Neuroscience* 2013;33:790. <https://doi.org/10.1523/JNEUROSCI.1238-12.2013>.
- [137] Liu S, Li Y, Choi HMC, Sarkar C, Koh EY, Wu J, et al. Lysosomal damage after spinal cord injury causes accumulation of RIPK1 and RIPK3 proteins and potentiation of necroptosis. *Cell Death & Disease* 2018;9. <https://doi.org/10.1038/S41419-018-0469-1>.

- [138] Hollis Ii ER. Axon Guidance Molecules and Neural Circuit Remodeling After Spinal Cord Injury. *Neurotherapeutics* 2016;13:360–9. <https://doi.org/10.1007/s13311-015-0416-0>.
- [139] Li H, Zheng J, Wang H, Becker ML, Leipzig ND. Neural stem cell encapsulation and differentiation in strain promoted crosslinked polyethylene glycol-based hydrogels. *J Biomater Appl* 2018;32:1222. <https://doi.org/10.1177/0885328218755711>.
- [140] Garbuzova-Davis S, Haller E, Navarro S, Besong TE, Boccio KJ, Hailu S, et al. Transplantation of Human Bone Marrow Stem Cells into Symptomatic ALS Mice Enhanced Structural and Functional Blood-Spinal Cord Barrier Repair HHS Public Access. *Exp Neurol* 2018;310:33–47. <https://doi.org/10.1016/j.expneurol.2018.08.012>.
- [141] Aumailley M. The laminin family. *Cell Adhesion & Migration* 2013;7:48. <https://doi.org/10.4161/CAM.22826>.
- [142] Baumann HJ, Mahajan G, Ham TR, Betonio P, Kothapalli CR, Shriver LP, et al. Softening of the chronic hemi-section spinal cord injury scar parallels dysregulation of cellular and extracellular matrix content. *J Mech Behav Biomed Mater* 2020;110:103953. <https://doi.org/10.1016/J.JMBBM.2020.103953>.
- [143] Schneider R, Bellenberg B, Gisevius B, Hirschberg S, Sankowski R, Prinz M, et al. Chitinase 3-like 1 and neurofilament light chain in CSF and CNS atrophy in MS. *Neurology(R) Neuroimmunology & Neuroinflammation* 2021;8. <https://doi.org/10.1212/NXI.0000000000000906>.
- [144] Gaur N, Perner C, Witte OW, Grosskreutz J. The Chitinases as Biomarkers for Amyotrophic Lateral Sclerosis: Signals From the CNS and Beyond. *Frontiers in Neurology* 2020;11:377. <https://doi.org/10.3389/FNEUR.2020.00377>.
- [145] Teshigawara K, Kuboyama T, Shigyo M, Nagata A, Sugimoto K, Matsuya Y, et al. A novel compound, denosomin, ameliorates spinal cord injury via axonal growth associated with astrocyte-secreted vimentin. *British Journal of Pharmacology* 2013;168:903. <https://doi.org/10.1111/J.1476-5381.2012.02211.X>.
- [146] Baldwin SA, Broderick R, Blades DA, Scheff3 SW. Alterations in Temporal/Spatial Distribution of GFAP-and Vimentin-Positive Astrocytes After Spinal Cord Contusion With the New York University Spinal Cord Injury Device. vol. 15. Mary Ann Liebert, Inc; 1998.

- [147] Redies C, Hertel N, Hübner CA. Cadherins and neuropsychiatric disorders. *Brain Research* 2012;1470:130–44. <https://doi.org/10.1016/J.BRAINRES.2012.06.020>.
- [148] Pancho A, Aerts T, Mitsogiannis MD, Seuntjens E. Protocadherins at the Crossroad of Signaling Pathways. *Frontiers in Molecular Neuroscience* 2020;13:117. <https://doi.org/10.3389/FNMOL.2020.00117/BIBTEX>.
- [149] Kim SY, Yasuda S, Tanaka H, Yamagata K, Kim H. Non-clustered protocadherin. *Cell Adhesion & Migration* 2011;5:97. <https://doi.org/10.4161/CAM.5.2.14374>.
- [150] Hara M, Kobayakawa K, Ohkawa Y, Kumamaru H, Yokota K, Saito T, et al. Interaction of reactive astrocytes with type i collagen induces astrocytic scar formation through the integrin-N-cadherin pathway after spinal cord injury. *Nature Medicine* 2017;23:818–28. <https://doi.org/10.1038/nm.4354>.
- [151] Rathore KI, Berard JL, Redensek A, Chierzi S, Lopez-Vales R, Santos M, et al. Lipocalin 2 Plays an Immunomodulatory Role and Has Detrimental Effects after Spinal Cord Injury. *The Journal of Neuroscience* 2011;31:13412. <https://doi.org/10.1523/JNEUROSCI.0116-11.2011>.
- [152] Smith JA, Braga A, Verheyen J, Basilico S, Bandiera S, Alfaro-Cervello C, et al. RNA Nanotherapeutics for the Amelioration of Astroglial Reactivity. *Molecular Therapy Nucleic Acids* 2018;10:103. <https://doi.org/10.1016/J.OMTN.2017.11.008>.
- [153] Decimo I, Bifari F, Rodriguez FJ, Malpeli G, Dolci S, Lavarini V, et al. Nestin- and Doublecortin-Positive Cells Reside in Adult Spinal Cord Meninges and Participate in Injury-Induced Parenchymal Reaction. *Stem Cells (Dayton, Ohio)* 2011;29:2062. <https://doi.org/10.1002/STEM.766>.
- [154] Wiese C, Rolletschek A, Kania G, Blyszczuk P, Tarasov K v., Tarasova Y, et al. Nestin expression - A property of multi-lineage progenitor cells? *Cellular and Molecular Life Sciences* 2004;61:2510–22. <https://doi.org/10.1007/s00018-004-4144-6>.
- [155] Yamamoto SI, Nagao M, Sugimori M, Kosako H, Nakatomi H, Yamamoto N, et al. Transcription Factor Expression and Notch-Dependent Regulation of Neural Progenitors in the Adult Rat Spinal Cord. *The Journal of Neuroscience* 2001;21:9814. <https://doi.org/10.1523/JNEUROSCI.21-24-09814.2001>.

- [156] Namiki J, Tator CH. Cell Proliferation and Nestin Expression in the Ependyma of the Adult Rat Spinal Cord after Injury. *Journal of Neuropathology & Experimental Neurology* 1999;58:489–98. <https://doi.org/10.1097/00005072-199905000-00008>.
- [157] Cawsey T, Duflou J, Weickert CS, Gorrie CA. Nestin-Positive Ependymal Cells Are Increased in the Human Spinal Cord after Traumatic Central Nervous System Injury. *Journal of Neurotrauma* 2015;32:1393. <https://doi.org/10.1089/NEU.2014.3575>.
- [158] Kozlova EN. Differentiation and migration of astrocytes in the spinal cord following dorsal root injury in the adult rat. *European Journal of Neuroscience* 2003;17:782–90. <https://doi.org/10.1046/j.1460-9568.2003.02518.x>.
- [159] Figueroa JD, Serrano-Illan M, Licero J, Cordero K, Miranda JD, de Leon M. Fatty Acid Binding Protein 5 Modulates Docosahexaenoic Acid-Induced Recovery in Rats Undergoing Spinal Cord Injury. *Journal of Neurotrauma* 2016;33:1436. <https://doi.org/10.1089/NEU.2015.4186>.
- [160] Cheng A, Jia W, Kawahata I, Fukunaga K. A novel fatty acid-binding protein 5 and 7 inhibitor ameliorates oligodendrocyte injury in multiple sclerosis mouse models. *EBioMedicine* 2021;72. <https://doi.org/10.1016/J.EBIOM.2021.103582>.
- [161] Peng X, Studholme K, Kanjiya MP, Luk J, Bogdan D, Elmes MW, et al. Fatty-acid-binding protein inhibition produces analgesic effects through peripheral and central mechanisms. *Molecular Pain* 2017;13. <https://doi.org/10.1177/1744806917697007>.
- [162] Wang GQ, Bonkovsky HL, de Lemos A, Burczynski FJ. Recent insights into the biological functions of liver fatty acid binding protein 1. *Journal of Lipid Research* 2015;56:2238. <https://doi.org/10.1194/JLR.R056705>.
- [163] Shin HY, Kim H, Kwon MJ, Hwang DH, Lee KY, Kim BG. Molecular and Cellular Changes in the Lumbar Spinal Cord following Thoracic Injury: Regulation by Treadmill Locomotor Training. *PLoS ONE* 2014;9. <https://doi.org/10.1371/JOURNAL.PONE.0088215>.
- [164] Du C, Wu H, Leng RP. UBE4B targets phosphorylated p53 at serines 15 and 392 for degradation. *Oncotarget* 2016;7:2823. <https://doi.org/10.18632/ONCOTARGET.6555>.
- [165] Dashzeveg N, Taira N, Lu ZG, Kimura J, Yoshida K. Palmdelphin, a novel target of p53 with Ser46 phosphorylation, controls cell death in response to DNA damage. *Cell Death and Disease* 2014;5. <https://doi.org/10.1038/cddis.2014.176>.

- [166] Martin LJ, Chen K, Liu Z. Adult Motor Neuron Apoptosis Is Mediated by Nitric Oxide and Fas Death Receptor Linked by DNA Damage and p53 Activation. *The Journal of Neuroscience* 2005;25:6449. <https://doi.org/10.1523/JNEUROSCI.0911-05.2005>.
- [167] Loughery J, Cox M, Smith LM, Meek DW. Critical role for p53-serine 15 phosphorylation in stimulating transactivation at p53-responsive promoters. *Nucleic Acids Research* 2014;42:7666. <https://doi.org/10.1093/NAR/GKU501>.
- [168] Smeenk L, van Heeringen SJ, Koeppel M, Gilbert B, Janssen-Megens E. Role of p53 Serine 46 in p53 Target Gene Regulation. *PLoS ONE* 2011;6:17574. <https://doi.org/10.1371/journal.pone.0017574>.
- [169] Castrogiovanni C, Waterschoot B, de Backer O, Dumont P. Serine 392 phosphorylation modulates p53 mitochondrial translocation and transcription-independent apoptosis. *Nature Publishing Group* 2017;25:190–203. <https://doi.org/10.1038/cdd.2017.143>.
- [170] Wu J, Yoo S, Wilcock D, Lytle JM, Leung PY, Colton CA, et al. Interaction of NG2+ glial progenitors and microglia/macrophages from the injured spinal cord. *Glia* 2010;58:410. <https://doi.org/10.1002/GLIA.20932>.
- [171] Buss A, Pech K, Kakulas BA, Martin D, Schoenen J, Noth J, et al. Matrix metalloproteinases and their inhibitors in human traumatic spinal cord injury. *BMC Neurology* 2007;7:17. <https://doi.org/10.1186/1471-2377-7-17>.
- [172] Tang J, Kang Y, Huang L, Wu L, Peng Y. TIMP1 preserves the blood–brain barrier through interacting with CD63/integrin  $\beta$ 1 complex and regulating downstream FAK/RhoA signaling. *Acta Pharmaceutica Sinica B* 2020;10:987. <https://doi.org/10.1016/J.APSB.2020.02.015>.
- [173] Shifman MI, Selzer ME. Differential Expression of Class 3 and 4 Semaphorins and Netrin in the Lamprey Spinal Cord during Regeneration. *J Comp Neurol* 2007;501:631. <https://doi.org/10.1002/CNE.21283>.
- [174] Wang F, Eagleson KL, Levitt P. Positive Regulation of Neocortical Synapse Formation by the Plexin-D1 Receptor. *Brain Res* 2015;1616:157. <https://doi.org/10.1016/J.BRAINRES.2015.05.005>.

- [175] Scott K, O'Rourke R, Gillen A, Appel B. Prdm8 regulates pMN progenitor specification for motor neuron and oligodendrocyte fates by modulating the Shh signaling response. *Development* 2020;147. <https://doi.org/10.1242/DEV.191023>.
- [176] Kearns CA, Walker M, Ravanelli AM, Scott K, Arzbecker MR, Appel B. Zebrafish spinal cord oligodendrocyte formation requires boc function. *Genetics* 2021;218. <https://doi.org/10.1093/GENETICS/IYAB082>.
- [177] Ogura T, Sakaguchi H, Miyamoto S, Takahashi J. Three-dimensional induction of dorsal, intermediate and ventral spinal cord tissues from human pluripotent stem cells. *Development (Cambridge)* 2018;145. <https://doi.org/10.1242/DEV.162214/VIDEO-1>.
- [178] Izumi H, Li Y, Shibaki M, Mori D, Yasunami M, Sato S, et al. Recycling endosomal CD133 functions as an inhibitor of autophagy at the pericentrosomal region. *Scientific Reports* 2019;9. <https://doi.org/10.1038/S41598-019-39229-8>.
- [179] Jászai J, Graupner S, Tanaka EM, Funk RHW, Huttner WB, Brand M, et al. Spatial Distribution of Prominin-1 (CD133) – Positive Cells within Germinative Zones of the Vertebrate Brain. *PLoS ONE* 2013;8. <https://doi.org/10.1371/JOURNAL.PONE.0063457>.
- [180] Luo Y, Coskun V, Liang A, Yu J, Cheng L, Ge W, et al. Single-Cell Transcriptome Analyses Reveal Signals to Activate Dormant Neural Stem Cells. *Cell* 2015;161:1175. <https://doi.org/10.1016/J.CELL.2015.04.001>.
- [181] Sasaki H, Ishikawa M, Tanaka N, Nakanishi K, Kamei N, Asahara T, et al. Administration of human peripheral blood-derived CD133+ cells accelerates functional recovery in a rat spinal cord injury model. *Spine (Phila Pa 1976)* 2009;34:249–54. <https://doi.org/10.1097/BRS.0B013E3181913CDE>.
- [182] Yang X, Tomita T, Wines-Samuelson M, Beglopoulos V, Tansey MG, Kopan R, et al. Notch1 signaling influences V2 interneuron and motor neuron development in the spinal cord. *Developmental Neuroscience* 2006;28:102–17. <https://doi.org/10.1159/000090757>.
- [183] Jalali A, Bassuk AG, Kan L, Israsena N, Mukhopadhyay A, McGuire T, et al. HeyL promotes neuronal differentiation of neural progenitor cells. *J Neurosci Res* 2011;89:299. <https://doi.org/10.1002/JNR.22562>.
- [184] Sobrido-Cameán D, Robledo D, Romaus-Sanjurjo D, Pérez-Cedrón V, Sánchez L, Rodicio MC, et al. Inhibition of Gamma-Secretase Promotes Axon Regeneration After a Complete

Spinal Cord Injury. *Frontiers in Cell and Developmental Biology* 2020;8.  
<https://doi.org/10.3389/FCELL.2020.00173/FULL>.

- [185] Duval N, Vaslin C, Barata TC, Frarma Y, Contremoulins V, Baudin X, et al. Bmp4 patterns smad activity and generates stereotyped cell fate organization in spinal organoids. *Development* (Cambridge) 2019;146.  
<https://doi.org/10.1242/DEV.175430/264854/AM/BMP4-PATTERNS-SMAD-ACTIVITY-AND-GENERATES>.
- [186] Farrukh F, Davies E, Berry M, Logan A, Ahmed Z. BMP4/Smad1 Signalling Promotes Spinal Dorsal Column Axon Regeneration and Functional Recovery After Injury. *Molecular Neurobiology* 2019;56:6807. <https://doi.org/10.1007/S12035-019-1555-9>.
- [187] Maxwell MM, Tomkinson EM, Nobles J, Wizeman JW, Amore AM, Quinti L, et al. The Sirtuin 2 microtubule deacetylase is an abundant neuronal protein that accumulates in the aging CNS. *Human Molecular Genetics* 2011;20:3986.  
<https://doi.org/10.1093/HMG/DDR326>.
- [188] González P, González-Fernández C, Javier Rodríguez F. Effects of Wnt5a overexpression in spinal cord injury. *Journal of Cellular and Molecular Medicine* 2021;25:5150.  
<https://doi.org/10.1111/JCMM.16507>.
- [189] Hiyama A, Yokoyama K, Nukaga T, Sakai D, Mochida J. A complex interaction between Wnt signaling and TNF- $\alpha$  in nucleus pulposus cells. *Arthritis Research & Therapy* 2013;15:R189. <https://doi.org/10.1186/AR4379>.
- [190] Krupnik VE, Sharp JD, Jiang C, Robison K, Chickering TW, Amaravadi L, et al. Functional and structural diversity of the human Dickkopf gene family. vol. 238. 1999.
- [191] Lemarchant S, Pruvost M, Montaner J, Emery E, Vivien D, Kanninen K, et al. ADAMTS proteoglycanases in the physiological and pathological central nervous system. *Journal of Neuroinflammation* 2013;10:133. <https://doi.org/10.1186/1742-2094-10-133>.
- [192] Post S, Weng Y-C, Cimprich K, Bo Chen L, Xu Y, Y-H Lee EP. Phosphorylation of serines 635 and 645 of human Rad17 is cell cycle regulated and is required for G 1 S checkpoint activation in response to DNA damage. 2001.

- [193] Bao S, Tibbetts RS, Brumbaugh KM, Fang Y, Richardson DA, Ali A, et al. ATR/ATM-mediated phosphorylation of human Rad17 is required for genotoxic stress responses. *Nature* 2001 411:6840 2001;411:969–74. <https://doi.org/10.1038/35082110>.
- [194] Nakamura H, Hoshino Y, Okuyama H, Matsuo Y, Yodoi J. Thioredoxin 1 delivery as new therapeutics. *Advanced Drug Delivery Reviews* 2009;61:303–9. <https://doi.org/10.1016/J.ADDR.2009.01.003>.
- [195] Guilcher SJT, Catharine Craven B, Bassett-Gunter RL, Cimino SR, Hitzig SL. An examination of objective social disconnectedness and perceived social isolation among persons with spinal cord injury/dysfunction: a descriptive cross-sectional study. *Disability and Rehabilitation* 2021;43:69–75. <https://doi.org/10.1080/09638288.2019.1616328>.
- [196] Onose G, Anghelescu A, Muresanu DF, Padure L, Haras MA, Co C, et al. REVIEW A review of published reports on neuroprotection in spinal cord injury. *Spinal Cord* 2009;47:716–26. <https://doi.org/10.1038/sc.2009.52>.
- [197] Onose G, Haras M, Anghelescu A, Mureşanu D, Giuglea C, Daia Chendreanu C, et al. Integrative emphases on intimate, intrinsic propensity/ pathological processes-causes of self recovery limits and also, subtle related targets for neuroprotection/ pleiotropicity/ multimodal actions, by accessible therapeutic approaches-in spinal cord injuries. vol. 3. n.d.
- [198] Onose G, Mureşanu DF, Ciurea A v, Daia Chendreanu\*’ \* C, Mihaescu AS, Mardare DC, et al. Neuroprotective and consequent neurorehabilitative clinical outcomes, in patients treated with the pleiotropic drug cerebrolysin. vol. 2. n.d.
- [199] Onose G, Haras M, Anghelescu A, Mureşanu D, Giuglea C, Daia Chendreanu C, et al. Integrative emphases on intimate, intrinsic propensity/ pathological processes-causes of self recovery limits and also, subtle related targets for neuroprotection/ pleiotropicity/ multimodal actions, by accessible therapeutic approaches-in spinal cord injuries. vol. 3. n.d.
- [200] Borgens RB, Shi R. Immediate recovery from spinal cord injury through molecular repair of nerve membranes with polyethylene glycol. vol. 14. 2000.
- [201] Luo J, Borgens R, Shi R. Polyethylene glycol immediately repairs neuronal membranes and inhibits free radical production after acute spinal cord injury. vol. 83. 2002.
- [202] Borgens RB, Bohnert D. Rapid Recovery From Spinal Cord Injury After Subcutaneously Administered Polyethylene Glycol. vol. 66. 2001.

- [203] Liu-Snyder P, Logan MP, Shi R, Smith DT, Borgens R ben. Neuroprotection from secondary injury by polyethylene glycol requires its internalization. *Journal of Experimental Biology* 2007;210:1455–62. <https://doi.org/10.1242/JEB.02756>.
- [204] Krause TL, Bittner GD. Rapid morphological fusion of severed myelinated axons by polyethylene glycol (nerve regeneration/axolemmal fusion/cytoplasmic repair/*Lumbricus terrestris*). vol. 87. 1990.
- [205] Laverty PH, Leskovar A, Breur GJ, Coates JR, Bergman RL, Widmer WR, et al. A Preliminary Study of Intravenous Surfactants in Paraplegic Dogs: Polymer Therapy in Canine Clinical SCI. *JOURNAL OF NEUROTRAUMA* 2004;21.
- [206] Rad I, Khodayari K, Alijanvand SH, Mobasheri H. Interaction of polyethylene glycol (PEG) with the membrane-binding domains following spinal cord injury (SCI): Introduction of a mechanism for SCI repair. *Journal of Drug Targeting* 2015;23:79–88. <https://doi.org/10.3109/1061186X.2014.956668>.
- [207] Ahuja CS, Nori S, Tetreault L, Wilson J, Kwon B, Harrop J, et al. Traumatic spinal cord injury - Repair and regeneration. *Clinical Neurosurgery* 2017;80:S22–90. <https://doi.org/10.1093/neuros/nyw080>.
- [208] Jiang X, Stockwell BR, Conrad M. Ferroptosis: mechanisms, biology and role in disease. *Nature Reviews Molecular Cell Biology* 2021;22:266–82. <https://doi.org/10.1038/s41580-020-00324-8>.
- [209] Vieira HLA, Alves PM, Vercelli A. Modulation of neuronal stem cell differentiation by hypoxia and reactive oxygen species. *Progress in Neurobiology* 2011;93:444–55. <https://doi.org/10.1016/J.PNEUROBIO.2011.01.007>.
- [210] Eyrich NW, Potts CR, Robinson MH, Maximov V, Kenney AM. Reactive Oxygen Species Signaling Promotes Hypoxia-Inducible Factor 1 Stabilization in Sonic Hedgehog-Driven Cerebellar Progenitor Cell Proliferation. 2019.
- [211] Sprick JD, Mallet RT, Przyklenk K, Rickards CA. Ischemic and Hypoxic Conditioning: Potential for Protection of Vital Organs. *Exp Physiol* 2019;104:278. <https://doi.org/10.1113/EP087122>.
- [212] Wu D, Yotnda P. Induction and Testing of Hypoxia in Cell Culture. *J Vis Exp* 2011:2899. <https://doi.org/10.3791/2899>.

- [213] Stoica SI, Bleotu C, Ciobanu V, Mirela Ionescu A, Albadi I, Onose G, et al. Considerations about Hypoxic Changes in Neuraxis Tissue Injuries and Recovery 2022. <https://doi.org/10.3390/biomedicines10020481>.
- [214] Gregg L Semenza. HIF-1 and mechanisms of hypoxia sensing 2001. [https://doi.org/doi:10.1016/s0955-0674\(00\)00194-0](https://doi.org/doi:10.1016/s0955-0674(00)00194-0).
- [215] Stoica SI, Bleotu C, Ciobanu V, Mirela Ionescu A, Albadi I, Onose G, et al. Considerations about Hypoxic Changes in Neuraxis Tissue Injuries and Recovery 2022. <https://doi.org/10.3390/biomedicines10020481>.
- [216] Stoica SI, Bleotu C, Ciobanu V, Mirela Ionescu A, Albadi I, Onose G, et al. Considerations about Hypoxic Changes in Neuraxis Tissue Injuries and Recovery 2022. <https://doi.org/10.3390/biomedicines10020481>.
- [217] Ratcliffe PJ. HIF-1 and HIF-2: Working alone or together in hypoxia? *Journal of Clinical Investigation* 2007;117:862–5. <https://doi.org/10.1172/JCI31750>.
- [218] Hewitson KS, McNeill LA, Riordan M v., Tian YM, Bullock AN, Welford RW, et al. Hypoxia-inducible factor (HIF) asparagine hydroxylase is identical to factor inhibiting HIF (FIH) and is related to the cupin structural family. *Journal of Biological Chemistry* 2002;277:26351–5. <https://doi.org/10.1074/JBC.C200273200>.
- [219] Stoica SI, Tănase I, Ciobanu V, Onose G. Initial researches on neuro-functional status and evolution in chronic ethanol consumers with recent traumatic spinal cord injury. *Journal of Medicine and Life* n.d.;12:97–112. <https://doi.org/10.25122/jml-2019-0026>.
- [220] Hao J, Chen X, Fu T, Liu J, Yu M, Han W, et al. The Expression of VHL (Von Hippel-Lindau) After Traumatic Spinal Cord Injury and Its Role in Neuronal Apoptosis. *Neurochemical Research* 2016;41:2391–400. <https://doi.org/10.1007/s11064-016-1952-7>.
- [221] Patrick H. Maxwell\* MSW-WCSCCECVECCCWWPERMPJR. The tumour suppressorprotein VHL targetshypoxia-inducible factors foroxygen-dependent proteolysis. *NATURE* 1999.
- [222] Wang GL, Jiang BH, Rue EA, Semenza GL. Hypoxia-inducible factor 1 is a basic-helix-loop-helix-PAS heterodimer regulated by cellular O<sub>2</sub> tension. *Proc Natl Acad Sci U S A* 1995;92:5510. <https://doi.org/10.1073/PNAS.92.12.5510>.

- [223] D'ignazio L, Rocha S. Hypoxia Induced NF- $\kappa$ B. *Cells* 2016;5:1–8. <https://doi.org/10.3390/CELLS5010010>.
- [224] Kerendi F, Halkos ME, Kin H, Corvera JS, Brat DJ, Wagner MB, et al. Upregulation of hypoxia inducible factor is associated with attenuation of neuronal injury in neonatal piglets undergoing deep hypothermic circulatory arrest. *Journal of Thoracic and Cardiovascular Surgery* 2005;130:1079.e1-1079.e10. <https://doi.org/10.1016/J.JTCVS.2005.05.045>.
- [225] Yu J, Yang H, Fang B, Zhang Z, Wang Y, Dai Y. mfat-1 transgene protects cultured adult neural stem cells against cobalt chloride-mediated hypoxic injury by activating Nrf2/ARE pathways. *J Neurosci Res* 2018;96:87–102. <https://doi.org/10.1002/JNR.24096>.
- [226] Magiorkinis G, Hurst TP, Miyazawa M, Brütting C, Narasimhan H, Hoffmann F, et al. Investigation of Endogenous Retrovirus Sequences in the Neighborhood of Genes Up-regulated in a Neuroblastoma Model after Treatment with Hypoxia-Mimetic Cobalt Chloride 2018. <https://doi.org/10.3389/fmicb.2018.00287>.
- [227] Zhong D, Cao Y, Li C-J, Li M, Rong Z-J, Jiang L, et al. Neural stem cell-derived exosomes facilitate spinal cord functional recovery after injury by promoting angiogenesis. *Experimental Biology and Medicine* 2020;245:54–65. <https://doi.org/10.1177/1535370219895491>.
- [228] Chouchay S, Noctor SC, Chutabhakdikul N. MICROGLIA ENHANCES PROLIFERATION OF NEURAL PROGENITOR CELLS IN AN IN VITRO MODEL OF HYPOXIC-ISCHEMIC INJURY. *EXCLI Journal* 2020;19:950–61. <https://doi.org/10.17179/excli2020-2249>.
- [229] Cameron NE, Cotter MA. Rapid Publication Neurovascular Dysfunction in Diabetic Rats Potential Contribution of Autoxidation and Free Radicals Examined Using Transition Metal Chelating Agents. vol. 96. 1995.
- [230] Kletkiewicz H, Klimiuk M, Woźniak AW, Mila-Kierzenkowska C, Dokładny K, Rogalska J. antioxidants How to Improve the Antioxidant Defense in Asphyxiated Newborns-Lessons from Animal Models n.d. <https://doi.org/10.3390/antiox9090898>.
- [231] Zhang R, Huang Q, Zou L, Cao X, Huang H, Chu X. Beneficial effects of deferoxamine against astrocyte death induced by modified oxygen glucose deprivation. *Brain Research* 2014;1583:23–33. <https://doi.org/10.1016/J.BRAINRES.2014.08.016>.

- [232] Llorente IL, Xie Y, Mazzitelli JA, Hatanaka EA, Cinkornpumin J, Miller DR, et al. Patient-derived glial enriched progenitors repair functional deficits due to white matter stroke and vascular dementia in rodents. *Science Translational Medicine* 2021;13. [https://doi.org/10.1126/SCITRANSLMED.AAZ6747/SUPPL\\_FILE/AAZ6747\\_SM.PDF](https://doi.org/10.1126/SCITRANSLMED.AAZ6747/SUPPL_FILE/AAZ6747_SM.PDF).
- [233] David BT, Curtin JJ, Brown JL, Coutts DJC, Boles NC, Hill CE. Treatment with hypoxia-mimetics protects cultured rat Schwann cells against oxidative stress-induced cell death. *Glia* 2021;69:2215–34. <https://doi.org/10.1002/GLIA.24019>.
- [234] Chu K, Jung KH, Kim SJ, Lee ST, Kim J, Park HK, et al. Transplantation of human neural stem cells protect against ischemia in a preventive mode via hypoxia-inducible factor-1alpha stabilization in the host brain. *Brain Res* 2008;1207:182–92. <https://doi.org/10.1016/J.BRAINRES.2008.02.043>.
- [235] Tang G, Chen Y, Chen J, Chen Z, Jiang W. Deferoxamine Ameliorates Compressed Spinal Cord Injury by Promoting Neovascularization in Rats. *Journal of Molecular Neuroscience* 2020;70:1437–44. <https://doi.org/10.1007/S12031-020-01564-1/FIGURES/6>.
- [236] Li G, Zhao Y, Li Y, Lu J. Up-Regulation of Neuronal Nitric Oxide Synthase Expression by Cobalt Chloride Through a HIF-1 $\alpha$  Mechanism in Neuroblastoma Cells. *Neuromolecular Med* 2015;17:443–53. <https://doi.org/10.1007/S12017-015-8373-7>.
- [237] Milosevic J, Adler I, Manaenko A, Schwarz SC, Walkinshaw G, Arend M, et al. Non-hypoxic stabilization of hypoxia-inducible factor alpha (HIF-alpha): relevance in neural progenitor/stem cells. *Neurotox Res* 2009;15:367–80. <https://doi.org/10.1007/S12640-009-9043-Z>.
- [238] Chen XH, Chen DT, Huang XM, Chen YH, Pan JH, Zheng XC, et al. Dexmedetomidine Protects Against Chemical Hypoxia-Induced Neurotoxicity in Differentiated PC12 Cells Via Inhibition of NADPH Oxidase 2-Mediated Oxidative Stress. *Neurotox Res* 2019;35:139–49. <https://doi.org/10.1007/S12640-018-9938-7>.
- [239] Yoo SY, Yoo JY, Kim HB, Baik TK, Lee JH, Woo RS. Neuregulin-1 Protects Neuronal Cells Against Damage due to CoCl<sub>2</sub>-Induced Hypoxia by Suppressing Hypoxia-Inducible Factor-1 $\alpha$  and P53 in SH-SY5Y Cells. *Int Neurourol J* 2019;23:S111–8. <https://doi.org/10.5213/INJ.1938190.095>.

- [240] LI F, ZHANG J, LIAO R, DUAN Y, TAO L, XU Y, et al. Mesenchymal stem cell-derived extracellular vesicles prevent neural stem cell hypoxia injury via promoting miR-210-3p expression. *Mol Med Rep* 2020;22:3813–21. <https://doi.org/10.3892/MMR.2020.11454>.
- [241] Li HL, Zaghoul N, Ahmed I, Omelchenko A, Firestein BL, Huang H, et al. Caffeine inhibits hypoxia-induced nuclear accumulation in HIF-1 $\alpha$  and promotes neonatal neuronal survival. *Exp Neurol* 2019;317:66–77. <https://doi.org/10.1016/J.EXPNEUROL.2019.01.014>.
- [242] Maliha AM, Kuehn S, Hurst J, Herms F, Fehr M, Bartz-Schmidt KU, et al. Diminished apoptosis in hypoxic porcine retina explant cultures through hypothermia. *Scientific Reports* 2019 9:1 2019;9:1–16. <https://doi.org/10.1038/s41598-019-41113-4>.
- [243] Crispo JAG, Ansell DR, Ubriaco G, Tai TC. Role of reactive oxygen species in the neural and hormonal regulation of the PNMT gene in PC12 cells. *Oxid Med Cell Longev* 2011;2011. <https://doi.org/10.1155/2011/756938>.
- [244] Li G, Zhao Y, Li Y, Lu J. Up-Regulation of Neuronal Nitric Oxide Synthase Expression by Cobalt Chloride Through a HIF-1 $\alpha$  Mechanism in Neuroblastoma Cells. *Neuromolecular Med* 2015;17:443–53. <https://doi.org/10.1007/S12017-015-8373-7>.
- [245] Merlo S, Luaces JP, Spampinato SF, Toro-Urrego N, Caruso GI, D'amico F, et al. SIRT1 Mediates Melatonin's Effects on Microglial Activation in Hypoxia: In Vitro and In Vivo Evidence. *Biomolecules* 2020;10. <https://doi.org/10.3390/BIOM10030364>.
- [246] Wenker SD, Chamorro ME, Vittori DC, Nesse AB. Protective action of erythropoietin on neuronal damage induced by activated microglia. *FEBS J* 2013;280:1630–42. <https://doi.org/10.1111/FEBS.12172>.
- [247] Wang P, Li L, Zhang Z, Kan Q, Chen S, Gao F. Time-dependent homeostasis between glucose uptake and consumption in astrocytes exposed to CoCl<sub>2</sub> treatment. *Mol Med Rep* 2016;13:2909–17. <https://doi.org/10.3892/MMR.2016.4873>.
- [248] Jeon ES, Shin JH, Hwang SJ, Moon GJ, Bang OY, Kim HH. Cobalt chloride induces neuronal differentiation of human mesenchymal stem cells through upregulation of microRNA-124a. *Biochem Biophys Res Commun* 2014;444:581–7. <https://doi.org/10.1016/J.BBRC.2014.01.114>.
- [249] C. Cernescu SR. *Practica diagnosticului virusologic*. Ed. Concept publishing; 1997.

- [250] <https://assets.thermofisher.com/TFS-Assets/BID/brochures/flow-cytometry-capabilities-guide-brochure.pdf> n.d.
- [251] <https://www.flowjo.com/solutions/flowjo> n.d.
- [252] McKinnon KM. Flow cytometry: An overview. *Current Protocols in Immunology* 2018;2018:5.1.1-5.1.11. <https://doi.org/10.1002/CPIM.40>.
- [253] Murray A. Cell cycle checkpoints. *Current Opinion in Cell Biology* 1994;6:872–6. [https://doi.org/10.1016/0955-0674\(94\)90059-0](https://doi.org/10.1016/0955-0674(94)90059-0).
- [254] Chomczynski P, Sacchi N. Single-Step Method of RNA Isolation by Acid Guanidinium Thiocyanate-Phenol-Chloroform Extraction. vol. 162. 1987.
- [255] [https://assets.fishersci.com/TFS-Assets/LSG/manuals/MAN0017977\\_highcap\\_cDNA\\_RT\\_UG.pdf](https://assets.fishersci.com/TFS-Assets/LSG/manuals/MAN0017977_highcap_cDNA_RT_UG.pdf) n.d.
- [256] Ma H, Bell KN, Loker RN. qPCR and qRT-PCR analysis: Regulatory points to consider when conducting biodistribution and vector shedding studies. *Molecular Therapy - Methods and Clinical Development* 2021;20:152–68. <https://doi.org/10.1016/J.OMTM.2020.11.007>.
- [257] <https://www.thermofisher.com/order/catalog/product/4448491?SID=srch-srp-4448491> n.d.
- [258] Schmittgen TD, Livak KJ. Analyzing real-time PCR data by the comparative CT method. *Nature Protocols* 2008;3:1101–8. <https://doi.org/10.1038/nprot.2008.73>.
- [259] [https://www.rndsystems.com/products/proteome-profiler-human-cell-stress-array-kit\\_ary018](https://www.rndsystems.com/products/proteome-profiler-human-cell-stress-array-kit_ary018) n.d.
- [260] [https://www.rndsystems.com/products/proteome-profiler-human-apoptosis-array-kit\\_ary009](https://www.rndsystems.com/products/proteome-profiler-human-apoptosis-array-kit_ary009) n.d.
- [261] (8400-0012-D00-Migration-Invasion-User-Manual3-Protocols-1--Data n. d. ). (8400-0012-D00-Migration-Invasion-User-Manual3-Protocols-1--Data, n.d.) n.d.
- [262] Takeoka A, Arber S. Functional Local Proprioceptive Feedback Circuits Initiate and Maintain Locomotor Recovery after Spinal Cord Injury. *Cell Reports* 2019;27:71-85.e3. <https://doi.org/10.1016/j.celrep.2019.03.010>.
- [263] <https://www.atcc.org/search#q=HTB%2011&sort=relevancy&numberOfResults=24> n.d.

- [264] Hari Y, Harashima N, Tajima Y, Harada M. Bcl-xL inhibition by molecular-targeting drugs sensitizes human pancreatic cancer cells to TRAIL. *Oncotarget* 2015;6:41902. <https://doi.org/10.18632/ONCOTARGET.5881>.
- [265] Jang C, Oh SF, Wada S, Rowe GC, Liu L, Chan MC, et al. A branched chain amino acid metabolite drives vascular transport of fat and causes insulin resistance HHS Public Access Author manuscript. *Nat Med* 2016;22:421–6. <https://doi.org/10.1038/nm.4057>.
- [266] Essen BioScience Inc. Scratch wound module User\_Manual. IncuCyte ZOOM® 96-Well Scratch Wound Cell Migration & Invasion Assays n.d. [https://qcri.queensu.ca/source/QCRI/Scratch%20wound%20module%20User\\_Manual.pdf](https://qcri.queensu.ca/source/QCRI/Scratch%20wound%20module%20User_Manual.pdf) (accessed June 19, 2022).
- [267] Ohri SS, Burke DA, Andres KR, Hetman M, Whittemore SR. Acute Neural and Proteostasis Messenger Ribonucleic Acid Levels Predict Chronic Locomotor Recovery after Contusive Spinal Cord Injury n.d. <https://doi.org/10.1089/neu.2020.7258>.
- [268] Halliday M, Hughes D, Mallucci GR. Fine-tuning PERK signaling for neuroprotection. *J Neurochem* 2017;142:812–26. <https://doi.org/10.1111/JNC.14112>.
- [269] Fernandez-Funez, Pedro. Cell Cycle Launching Hsp70 neuroprotection 2014. <https://doi.org/10.4161/cc.29148>.
- [270] <https://www.atcc.org/search#q=HTB%2014&sort=relevancy&numberOfResults=24> n.d.
- [271] [https://ec.europa.eu/environment/chemicals/lab\\_animals/legislation\\_en.htm](https://ec.europa.eu/environment/chemicals/lab_animals/legislation_en.htm) n.d.
- [272] Mizuno T, Goto Y, Baba K, Masuda K, Ohno K, Tsujimoto H. Molecular cloning of feline tumour necrosis factor receptor type I (TNFR I) and expression of TNFR I and TNFR II in lymphoid cells in cats. vol. 30. 2003.
- [273] Pennica DN; HJS; SPH; DRPMA; KWJ; ABB; GD v. Human tumour necrosis factor: precursor structure, expression and homology to lymphotoxin. *Nature* 1984.
- [274] Stoica SI, Onose G, Hotetiu M, Munteanu C. Effects of ethanol and deferoxamine on rat primary glial cell cultures, in regard with ischemia induced by traumatic spinal cord injury. *Balneo and PRM Research Journal* 2022:502. <https://doi.org/10.12680/balneo.2022.502>.
- [275] Sengupta P. The Laboratory Rat: Relating Its Age with Human's. vol. 4. 2013.

- [276] Foo LC, Allen NJ, Bushong EA, Ventura PB, Chung WS, Zhou L, et al. Development of a method for the purification and culture of rodent astrocytes. *Neuron* 2011;71:799–811. <https://doi.org/10.1016/J.NEURON.2011.07.022>.
- [277] kolajaan. NB: Unofficial translation Legally binding texts are those in Finnish and Swedish Act on the Protection of Animals Used for Scientific or Educational Purposes (497/2013). n.d.
- [278] Handbook of WMA Policies World Medical Association | WMA STATEMENT ON ANIMAL USE IN BIOMEDICAL RESEARCH. n.d.
- [279] Sarc L, Wraber B, Lipnik-Stangelj M. Ethanol and acetaldehyde disturb TNF-alpha and IL-6 production in cultured astrocytes n.d. <https://doi.org/10.1177/0960327110388533>.
- [280] Luczak A, Kubo Y. Predictive Neuronal Adaptation as a Basis for Consciousness. *Frontiers in Systems Neuroscience* 2022;15. <https://doi.org/10.3389/fnsys.2021.767461>.
- [281] Lebedev M, Buffo A, Mateos-Aparicio P, Rodríguez-Moreno A. The Impact of Studying Brain Plasticity. *Frontiers in Cellular Neuroscience* | *WwwFrontiersinOrg* 2019;13:66. <https://doi.org/10.3389/fncel.2019.00066>.
- [282] Walker JR, Detloff MR. Plasticity in cervical motor circuits following spinal cord injury and rehabilitation. *Biology (Basel)* 2021;10. <https://doi.org/10.3390/biology10100976>.
- [283] Abrahao KP, Salinas AG, Lovinger DM. Alcohol and the Brain: Neuronal Molecular Targets, Synapses, and Circuits. *Neuron* 2017;96:1223–38. <https://doi.org/10.1016/j.neuron.2017.10.032>.
- [284] Tateno M, Saito T. Biological Studies on Alcohol-Induced Neuronal Damage Correspondence n.d.
- [285] Abrahao KP, Salinas AG, Lovinger DM. Alcohol and the Brain: Neuronal Molecular Targets, Synapses, and Circuits. *Neuron* 2017;96:1223–38. <https://doi.org/10.1016/j.neuron.2017.10.032>.
- [286] Antonio AM, Druse MJ. Antioxidants Prevent Ethanol-Associated Apoptosis in Fetal Rhombencephalic Neurons. *Brain Res* 2008;1204:16–23. <https://doi.org/10.1016/j.brainres.2008.02.018>.

- [287] Antonio AM, Druse MJ. Antioxidants prevent ethanol-associated apoptosis in fetal rhombencephalic neurons. *Brain Research* 2008;1204:16–23. <https://doi.org/10.1016/j.brainres.2008.02.018>.
- [288] PubChem Compound Summary for CID 702, Ethanol. National Center for Biotechnology Information 2022. <https://pubchem.ncbi.nlm.nih.gov/compound/702> (accessed August 4, 2022).
- [289] Yukitake H, Takizawa M, Kimura H. Macrophage Migration Inhibitory Factor as an Emerging Drug Target to Regulate Antioxidant Response Element System 2017. <https://doi.org/10.1155/2017/8584930>.
- [290] Cox CS, McKay SE, Holmbeck MA, Christian BE, Scortea AC, Tsay AJ, et al. Mitohormesis in Mice via Sustained Basal Activation of Mitochondrial and Antioxidant Signaling. *Cell Metabolism* 2018;28:776-786.e5. <https://doi.org/10.1016/J.CMET.2018.07.011>.
- [291] Wallenborg K, Vlachos P, Eriksson S, Huijbregts L, Arnér ESJ, Joseph B, et al. Red wine triggers cell death and thioredoxin reductase inhibition: Effects beyond resveratrol and SIRT1. *Experimental Cell Research* 2009;315:1360–71. <https://doi.org/10.1016/j.yexcr.2009.02.022>.
- [292] MacHicao F, Muresanu DF, Hundesberger H, Pflüger M, Guekht A. Pleiotropic neuroprotective and metabolic effects of Actovegin's mode of action. *Journal of the Neurological Sciences* 2012;322:222–7. <https://doi.org/10.1016/j.jns.2012.07.069>.
- [293] de Decker S, Moore SA, Tipold A, Olby NJ, Stein V, Granger N. Current Approaches to the Management of Acute Thoracolumbar Disc Extrusion in Dogs. *Frontiers in Veterinary Science | WwwFrontiersinOrg* 2020;1:610. <https://doi.org/10.3389/fvets.2020.00610>.